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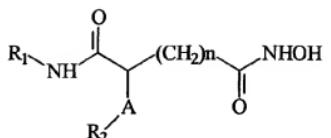
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(54) Title: NOVEL CLASS OF CYTODIFFERENTIATING AGENTS AND HISTONE DEACETYLASE INHIBITORS, AND METHODS OF USE THEREOF



(1)

(57) Abstract: The present invention provides the compound having formula (I), wherein each of R_1 and R_2 is, substituted or unsubstituted, aryl, cycloalkyl, cycloalkylamino, naphthyl, pyridineaminyl, piperidinyl, t-butyl, arylxoy, arylalkyloxy, or pyridine group; wherein A is an amide moiety, $-O-$, $-S-$, $-NH-$, or $-CH_2-$; and wherein n is an integer from 3 to 8. The present invention also provides a method of selectively inducing growth arrest, terminal differentiation and/or apoptosis of neoplastic cells and thereby inhibiting proliferation of such cells. Moreover, the present invention provides a method of treating a patient having a tumor characterized by proliferation of neoplastic cells. Lastly, the present invention provides a pharmaceutical composition comprising a pharmaceutically acceptable carrier and a therapeutically acceptable amount of the compound above.

WO 01/18171 A2

5 NOVEL CLASS OF CYTODIFFERENTIATING AGENTS AND
HISTONE DEACETYLASE INHIBITORS, AND METHODS OF USE THEREOF

This application claims the benefit of U.S. Provisional Application No. 60/208,688, filed June 1, 2000, and U.S. Provisional Application No. 60/152,755, filed September 8, 1999.

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Throughout this application various publications are referenced by Arabic numerals within parentheses. Full citations for these publications may be found at the end of the specification immediately preceding the claims. The disclosures of these 15 publications in their entireties are hereby incorporated by reference into this application in order to more fully describe the state of the art to which this invention pertains.

Background of the Invention

20 Cancer is a disorder in which a population of cells has become, in varying degrees, unresponsive to the control mechanisms which normally govern proliferation and differentiation. A recent approach to cancer therapy has been to attempt induction of terminal differentiation of the neoplastic cells (1). In cell 25 culture models differentiation has been reported by exposure of cells to a variety of stimuli, including: cyclic AMP and retinoic acid (2,3), aclarubicin and other anthracyclines (4).

There is abundant evidence that neoplastic transformation does 30 not necessarily destroy the potential of cancer cells to differentiate (1,5,6). There are many examples of tumor cells which do not respond to the normal regulators of proliferation and appear to be blocked in the expression of their differentiation program, and yet can be induced to differentiate 35 and cease replicating. A variety of agents, including some relatively simple polar compounds (5,7-9), derivatives of vitamin D and retinoic acid (10-12), steroid hormones (13), growth factors (6,14), proteases (15,16), tumor promoters

(17,18), and inhibitors of DNA or RNA synthesis (4,19-24), can induce various transformed cell lines and primary human tumor explants to express more differentiated characteristics.

5 Early studies by the some of present inventors identified a series of polar compounds that were effective inducers of differentiation in a number of transformed cell lines (8,9). One such effective inducer was the hybrid polar/apolar compound N,N'-hexamethylene bisacetamide (HMBA) (9), another was 10 suberoylanilide hydroxamic acid (SAHA) (39, 50). The use of these compounds to induce murine erythroleukemia (MEL) cells to undergo erythroid differentiation with suppression of oncogenicity has proved a useful model to study inducer-mediated differentiation of transformed cells (5,7-9).

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HMBA-induced MEL cell terminal erythroid differentiation is a multistep process. Upon addition of HMBA to MEL cells (745A-DS19) in culture, there is a latent period of 10 to 12 hours before commitment to terminal differentiation is detected. 20 Commitment is defined as the capacity of cells to express terminal differentiation despite removal of inducer (25). Upon continued exposure to HMBA there is progressive recruitment of cells to differentiate. The present inventors have reported that MEL cell lines made resistant to relatively low levels of 25 vincristine become markedly more sensitive to the inducing action of HMBA and can be induced to differentiate with little or no latent period (26).

HMBA is capable of inducing phenotypic changes consistent with 30 differentiation in a broad variety of cells lines (5). The characteristics of the drug induced effect have been most extensively studied in the murine erythroleukemia cell system (5,25,27,28). MEL cell induction of differentiation is both time and concentration dependent. The minimum concentration 35 required to demonstrate an effect in vitro in most strains is 2 to 3 mM; the minimum duration of continuous exposure generally

required to induce differentiation in a substantial portion (>20%) of the population without continuing drug exposure is about 36 hours.

5 There is evidence that protein kinase C is involved in the pathway of inducer-mediated differentiation (29). The *in vitro* studies provided a basis for evaluating the potential of HMBA as a cytodifferentiation agent in the treatment of human cancers (30). Several phase I clinical trials with HMBA have been 10 completed (31-36). Clinical trials have shown that this compound can induce a therapeutic response in patients with cancer (35,36). However, these phase I clinical trials also have demonstrated that the potential efficacy of HMBA is limited, in part, by dose-related toxicity which prevents 15 achieving optimal blood levels and by the need for intravenous administration of large quantities of the agent, over prolonged periods. Thus, some of the present inventors have turned to synthesizing compounds that are more potent and possibly less toxic than HMBA (37).

20

Recently, a class of compounds that induce differentiation, have been shown to inhibit histone deacetylases. Several experimental antitumor compounds, such as trichostatin A (TSA), trapoxin, suberoylanilide hydroxamic acid (SAHA), and 25 phenylbutyrate have been shown to act, at least in part, by inhibiting histone deacetylases (38, 39, 42). Additionally, diallyl sulfide and related molecules (43), oxamflatin, (44), MS-27-275, a synthetic benzamide derivative, (45) butyrate derivatives (46), FR901228 (47), depudecin (48), and m- 30 carboxy cinnamic acid bishydroxamide (39) have been shown to inhibit histone deacetylases. *In vitro*, these compounds can inhibit the growth of fibroblast cells by causing cell cycle arrest in the G1 and G2 phases (49-52), and can lead to the terminal differentiation and loss of transforming potential of 35 a variety of transformed cell lines (49-51). *In vivo*, phenylbutyrate is effective in the treatment of acute

promyelocytic leukemia in conjunction with retinoic acid (53). SAHA is effective in preventing the formation of mammary tumors in rats, and lung tumors in mice (54, 55).

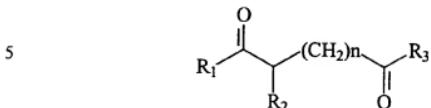
5 U.S. Patent No. 5,369,108 (41) issued to some of the present inventors discloses compounds useful for selectively inducing terminal differentiation of neoplastic cells, which compounds have two polar end groups separated by a flexible chain of 10 methylene groups, wherein one or both of the polar end groups 10 is a large hydrophobic group. Such compounds are stated to be more active than HMBA and HMBA related compounds.

However, U.S. Patent No. 5,369,108 does not disclose that an 15 additional large hydrophobic group at the same end of the molecule as the first hydrophobic group would further increase differentiation activity about 100 fold in an enzymatic assay and about 50 fold in a cell differentiation assay.

This new class of compounds of the present invention may be 20 useful for selectively inducing terminal differentiation of neoplastic cells and therefore aid in treatment of tumors in patients.

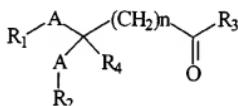
Summary of the Invention

The subject invention provides a compound having the formula:



wherein R_1 and R_2 are the same or different and are each a hydrophobic moiety; wherein R_3 is hydroxamic acid, hydroxylamino, hydroxyl, amino, alkylamino, or alkyloxy group; and n is an integer from 3 to 10, or a pharmaceutically acceptable salt thereof.

The subject invention also provides A compound having the 15 formula:



20 wherein each of R_1 and R_2 is, substituted or unsubstituted, aryl, cycloalkyl, cycloalkylamino, naphtha, pyridineamino, piperidino, 9-purine-6-amine, thiazoleamino group, hydroxyl, branched or unbranched alkyl, alkenyl, alkyloxy, aryloxy, arylalkyloxy, or pyridine group; wherein R_3 is hydroxamic acid, hydroxylamino, 25 hydroxyl, amino, alkylamino, or alkyloxy group; wherein R_4 is hydrogen, a halogen, a phenyl, or a cycloalkyl moiety; wherein A may be the same or different and represents an amide moiety, $-\text{O}-$, $-\text{S}-$, $-\text{NR}_5-$, or $-\text{CH}_2-$, where R_5 is a substituted or unsubstituted C_1-C_5 alkyl; and wherein n is an integer from 3 to 30 10, or a pharmaceutically acceptable salt thereof.

The subject invention also provides a method of selectively inducing terminal differentiation of neoplastic cells and thereby inhibiting proliferation of such cells which comprises 35 contacting the cells under suitable conditions with an effective amount of the aforementioned compound.

Description of the Figures

Figure 1. The effect of Compound 1 according to the subject invention on MEL cell differentiation.

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Figure 2. The effect of Compound 1 according to the subject invention on Histone Deacetylase 1 activity.

Figure 3. The effect of Compound 2 according to the subject invention on MEL cell differentiation.

Figure 4. The effect of Compound 3 according to the subject invention on MEL cell differentiation.

15 **Figure 5.** The effect of Compound 3 according to the subject invention on Histone Deacetylase 1 activity.

Figure 6. The effect of Compound 4 according to the subject invention on MEL cell differentiation.

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Figure 7. The effect of Compound 4 according to the subject invention on Histone Deacetylase 1 activity.

25 1 **Figure 8.** A photoaffinity label (3H-498) binds directly to HDAC

Figure 9. SAHA causes accumulation of acetylated histones H3 and H4 in the CWR22 tumor xenograft in mice.

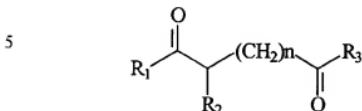
30 **Figure 10.** SAHA causes accumulation of acetylation histones H3 and H4 in peripheral blood mononuclear cells in patients. SAHA was administered by IV infusion daily x 3. Samples were isolated before (Pre), following infusion (Post) and 2 hours after infusion.

35

Figures 11a-11f. Show the effect of selected compounds on affinity purified human epitope-tagged (Flag) HDAC1.

Detailed Description of the Invention

The subject invention provides a compound having the formula:



wherein R_1 and R_2 are the same or different and are each a 10 hydrophobic moiety; wherein R_3 is hydroxamic acid, hydroxylamino, hydroxyl, amino, alkylamino, or alkyloxy group; and n is an integer from 3 to 10; or a pharmaceutically acceptable salt of the compound.

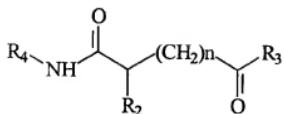
15 In the foregoing compound each of R_1 and R_2 is directly attached or through a linker, and is, substituted or unsubstituted, aryl, cycloalkyl, cycloalkylamino, naphtha, pyridineamino, piperidino, 9-purine-6-amine, thiazoleamino group, hydroxyl, branched or unbranched alkyl, alkenyl, alkyloxy, aryloxy, arylalkyloxy, or 20 pyridine group.

Where a linker is used, the linker may be an amide moiety, $-\text{O}-$, $-\text{S}-$, $-\text{NH}-$, or $-\text{CH}_2-$.

25 According to this invention, n may be 3-10, preferably 3-8, more preferably 3-7, yet more preferably 4, 5 or 6, and most preferably 5.

In another embodiment of the invention, the compound has the formula:

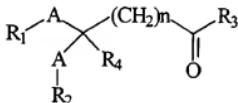
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wherein each of R₄ is, substituted or unsubstituted, aryl, 10 cycloalkyl, cycloalkylamino, naphtha, pyridineamino, piperidino, 9-purine-6-amine, thiazoleamino group, hydroxyl, branched or unbranched alkyl, alkenyl, alkyloxy, aryloxy, arylalkyloxy, or pyridine group. R₂ may be -amide-R₅, wherein R₅ is, substituted or unsubstituted, aryl, cycloalkyl, cycloalkylamino, naphtha, 15 pyridineamino, piperidino, 9-purine-6-amine, thiazoleamino group, hydroxyl, branched or unbranched alkyl, alkenyl, alkyloxy, aryloxy, arylalkyloxy, or pyridine group.

In a further embodiment of the invention the compound has the 20 formula:

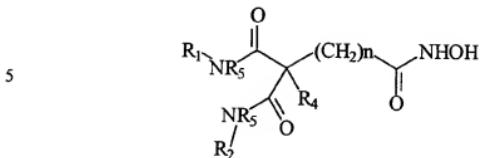
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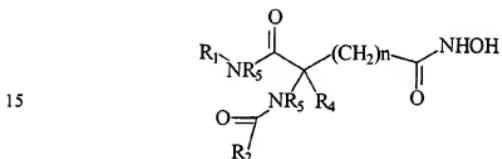
wherein each of R₁ and R₂ is, substituted or unsubstituted, aryl, cycloalkyl, cycloalkylamino, naphtha, pyridineamino, piperidino, 9-purine-6-amine, thiazoleamino group, hydroxyl, branched or unbranched alkyl, alkenyl, alkyloxy, aryloxy, arylalkyloxy, or 30 pyridine group; wherein R₃ is hydroxamic acid, hydroxylamino, hydroxyl, amino, alkyloamino, or alkyloxy group; wherein R₄ is hydrogen, a halogen, a phenyl, or a cycolalkyl moiety; wherein A may be the same or different and represents an amide moiety, -O-, -S-, -NR₅-, or -CH₂-, where R₅ is a substituted or 35 unsubstituted C₁-C₅ alkyl; and wherein n is an integer from 3 to 10, or a pharmaceutically acceptable salt thereof.

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In another embodiment the compound has the formula:

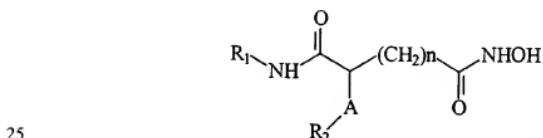


10 In yet another embodiment, the compound has the formula:



In a further embodiment, the compound has the formula:

20



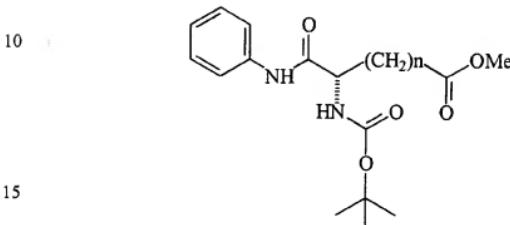
wherein each of R₁ and R₂ is, substituted or unsubstituted, aryl, cycloalkyl, cycloalkylamino, naphtha, pyridineamino, piperidino, t-butyl, aryloxy, arylalkyloxy, or pyridine group; and wherein 30 n is an integer from 3 to 8.

The aryl or cycloalkyl group may be substituted with a methyl, cyano, nitro, trifluoromethyl, amino, aminocarbonyl, methylcyano, chloro, fluoro, bromo, iodo, 2,3-difluoro, 2,4-difluoro, 2,5-difluoro, 3,4-difluoro, 3,5-difluoro, 2,6-difluoro, 1,2,3-trifluoro, 2,3,6-trifluoro, 2,4,6-trifluoro,

-11-

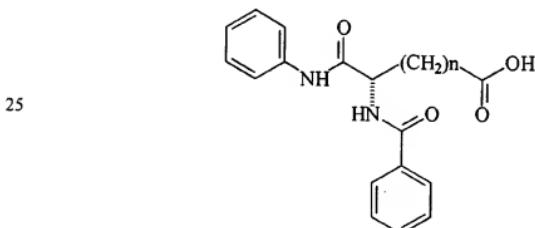
3,4,5-trifluoro, 2,3,5,6-tetrafluoro, 2,3,4,5,6-pentafluoro, azido, hexyl, t-butyl, phenyl, carboxyl, hydroxyl, methoxy, phenoxy, benzyloxy, phenylaminoxy, phenylaminocarbonyl, methyoxy carbonyl, methylaminocarbonyl, dimethylamino, 5 dimethylaminocarbonyl, or hydroxylaminocarbonyl group.

In a further embodiment, the compound has the formula:



or an enantiomer thereof.

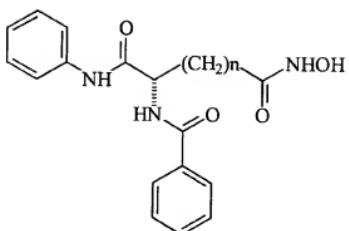
20 In a yet further embodiment, the compound has the formula:



or an enantiomer thereof.

In a further embodiment, the compound has the formula:

5



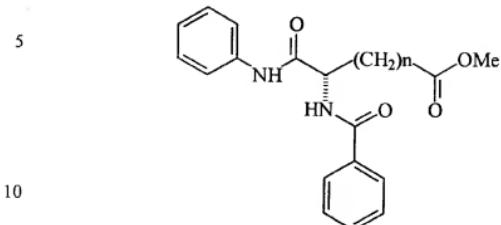
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or an enantiomer thereof.

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In a yet further embodiment, the compound has the formula:

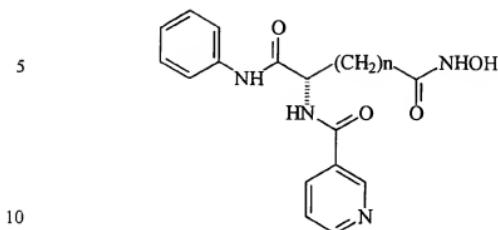


or an enantiomer thereof.

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-14-

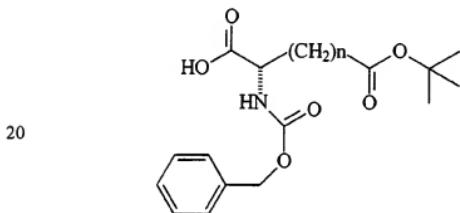
In a further embodiment, the compound has the formula:



or an enantiomer thereof.

In a yet further embodiment, the compound has the formula:

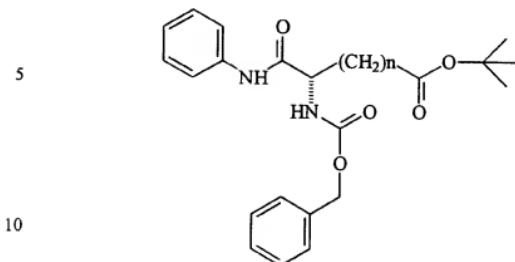
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25 or an enantiomer thereof.

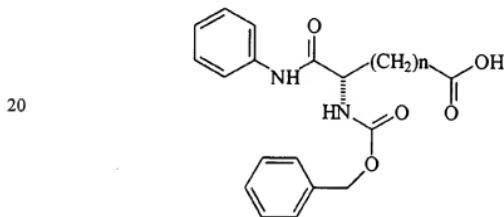
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In a yet further embodiment, the compound has the formula:



or an enantiomer thereof.

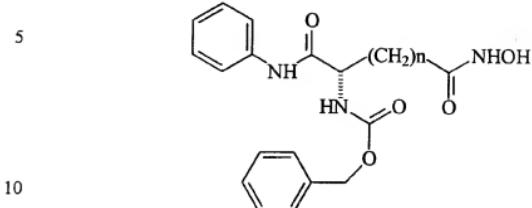
15 In a further embodiment, the compound has the formula:



or an enantiomer thereof.

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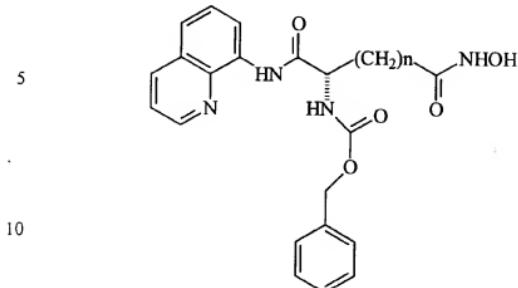
In a further embodiment, the compound has the formula:



or an enantiomer thereof.

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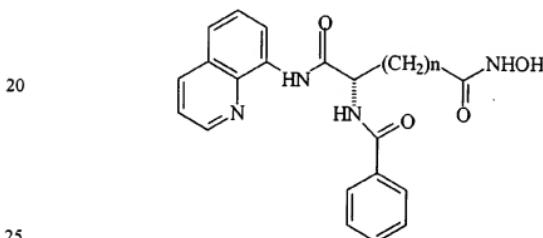
In a yet further embodiment, the compound has the formula:



or an enantiomer thereof.

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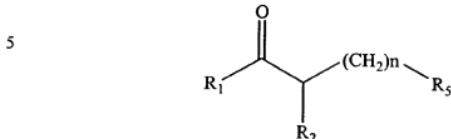
In a further embodiment, the compound has the formula:



or an enantiomer thereof.

This invention is also intended to encompass enantiomers and
30 salts of the compounds listed above.

In a further embodiment, the compound has the formula:



wherein R₁ and R₂ are the same or different and are each a hydrophobic moiety;

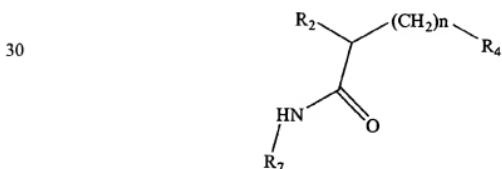
wherein R₅ is -C(O)-NHOH (hydroxamic acid), -C(O)-CF₃ (trifluoroacetyl), -NH-P(O)OH-CH₃, -SO₂NH₂ (sulfonamide), -SH (thiol), -C(O)-R₆, wherein R₆ is hydroxyl, amino, alkylamino, or alkyloxy group; and

15 n is an integer from 3 to 10, or a pharmaceutically acceptable salt thereof.

In the foregoing compound, each of R₁ and R₂ may be directly attached or through a linker, and is, substituted or 20 unsubstituted, aryl, cycloalkyl, cycloalkylamino, naphtha, pyridineamino, piperidino, 9-purine-6-amine, thiazoleamino group, hydroxyl, branched or unbranched alkyl, alkenyl, alkyloxy, aryloxy, arylalkyloxy, or pyridine group.

25 The linker may be an amide moiety, -O-, -S-, -NH-, or -CH₂-.

In another embodiment, the compound has the formula:



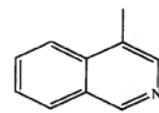
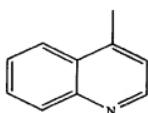
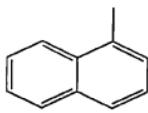
35 wherein each of R_i is, substituted or unsubstituted, aryl, cycloalkyl, cycloalkylamino, naphtha, pyridineamino, piperidino,

9-purine-6-amine, thiazoleamino group, hydroxyl, branched or unbranched alkyl, alkenyl, alkyloxy, aryloxy, arylalkyloxy, or pyridine group.

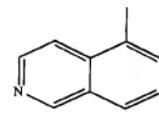
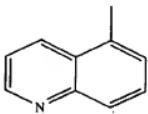
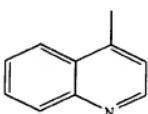
5 In the foregoing compound, R₂ may be -sulfonamide-R₈, or -amide-R₈, wherein R₈ is, substituted or unsubstituted, aryl, cycloalkyl, cycloalkylamino, naphtha, pyridineamino, piperidino, 9-purine-6-amine, thiazoleamino group, hydroxyl, branched or unbranched alkyl, alkenyl, alkyloxy, aryloxy, arylalkyloxy, or 10 pyridine group.

The R₂ may be -NH-C(O)-Y, -NH-SO₂-Y, wherein Y is selected from the group consisting of:

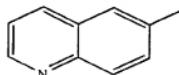
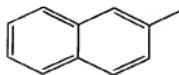
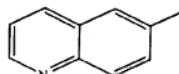
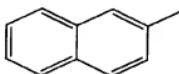
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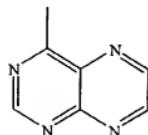
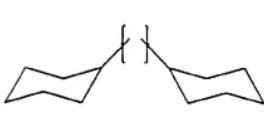
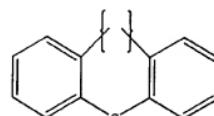
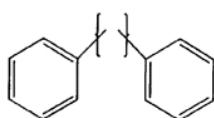
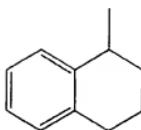
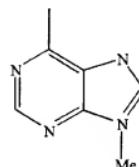
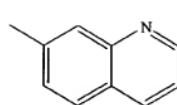
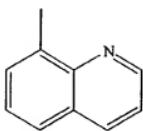


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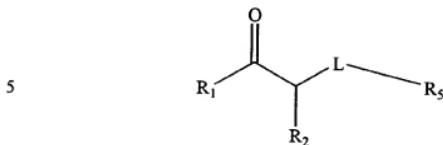


30 .

The R₇ may be selected from the group consisting of:



In yet another embodiment, the compound has the formula:



wherein R₁ and R₂ are the same or different and are each a hydrophobic moiety;

10 wherein R₅ is -C(O)-NHOH (hydroxamic acid), -C(O)-CF₃ (trifluoroacetyl), -NH-P(O)OH-CH₃, -SO₂NH₂ (sulfonamide), -SH (thiol), -C(O)-R₆, wherein R₆ is hydroxyl, amino, alkylamino, or alkyloxy group; and

15 wherein L is a linker consisting of -(CH₂)-, -C(O)-, -S-, -O-, -(CH=CH)-, -phenyl-, or -cycloalkyl-, or any combination thereof,

or a pharmaceutically acceptable salt thereof.

L may also be a linker consisting of -(CH₂)_n-, -C(O)-, -S-, -O-, 20 -(CH=CH)_m-, -phenyl-, or -cycloalkyl-, or any combination thereof, wherein n is an integer from 3 to 10, and m is an integer from 0 to 10,

In the foregoing compound, n may be from 4-7, and m is from 0-7. 25 Preferably n is 5 or 6, most preferably n is 6. Preferably m is from 1-6, more preferably m is 2-5, most preferably m is 3 or 4,

In the compound, each of R₁ and R₂ may be directly attached or 30 through a linker, and is, substituted or unsubstituted, aryl, cycloalkyl, cycloalkylamino, naphtha, pyridineamino, piperidino, 9-purine-6-amine, thiazoleamino group, hydroxyl, branched or unbranched alkyl, alkenyl, alkyloxy, aryloxy, arylalkyloxy, or pyridine group.

35

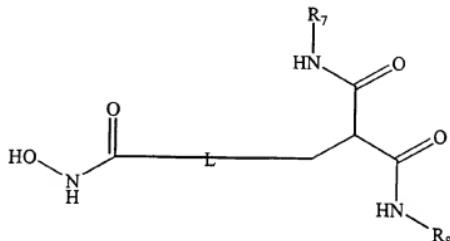
The linker may be an amide moiety, -O-, -S-, -NH-, or -CH₂-.

This invention is also intended to encompass enantiomers, salts and pro-drugs of the compounds disclosed herein.

In another embodiment the compound may have the formula:

5

10

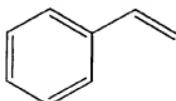


15 wherein L is a linker selected from the group consisting of -
 $(CH_2)-$, - $(CH=CH)-$, -phenyl-, -cycloalkyl-, or any combination thereof; and

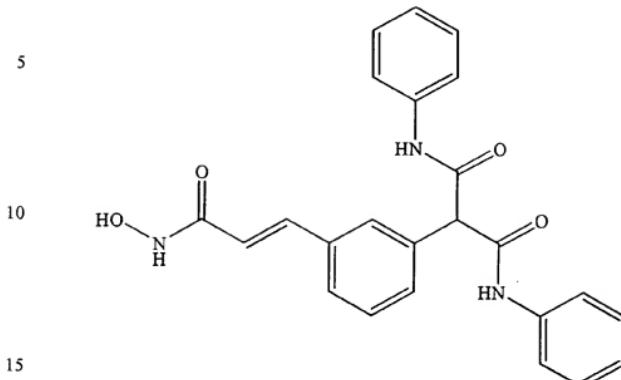
wherein each of R_7 and R_8 are independently substituted or
 20 unsubstituted, aryl, cycloalkyl, cycloalkylamino, naphtha, pyridineamino, piperidino, 9-purine-6-amine, thiazoleamino group, hydroxyl, branched or unbranched alkyl, alkenyl, alkyloxy, aryloxy, arylalkyloxy, or pyridine group.

In a preferred embodiment, the linker L comprises the moiety
 25

30



In another preferred embodiment, the compound has the formula:



Any of the disclosed compounds can be formed into a pharmaceutical composition together with a pharmaceutically acceptable carrier.

20

Any of the compounds can also be formed into a pharmaceutically acceptable salt of the compound using well known pharmacological techniques.

25 A prodrug of any of the compounds can also be made using well known pharmacological techniques.

Any of the compounds can be used in a method of inducing differentiation of tumor cells in a tumor comprising contacting 30 the cells with an effective amount of the compound so as to thereby differentiate the tumor cells.

Any of the compounds can also be used in a method of inhibiting the activity of histone deacetylase comprising contacting the 35 histone deacetylase with an effective amount of the compound so as to thereby inhibit the activity of histone deacetylase.

This invention, in addition to the above listed compounds, is further intended to encompass the use of homologs and analogs of such compounds. In this context, homologs are molecules having substantial structural similarities to the 5 above-described compounds and analogs are molecules having substantial biological similarities regardless of structural similarities.

In a further embodiment, the subject invention provides a 10 pharmaceutical composition comprising a pharmaceutically effective amount of any one of the aforementioned compounds and a pharmaceutically acceptable carrier.

In a yet further embodiment, the subject invention provides a 15 method of selectively inducing growth arrest, terminal differentiation and/or apoptosis of neoplastic cells and thereby inhibiting proliferation of such cells which comprises contacting the cells under suitable conditions with an effective amount of any one of the aforementioned compounds.

20

The contacting should be performed continuously for a prolonged period of time, i.e. for at least 48 hours, preferably for about 4-5 days or longer.

25 The method may be practiced *in vivo* or *in vitro*. If the method is practiced *in vitro*, contacting may be effected by incubating the cells with the compound. The concentration of the compound in contact with the cells should be from about 1 nM to about 25 mM, preferably from about 20 nM to about 25 mM, more preferably 30 from about 40 nM to 100 μ M, yet more preferably from about 40 nM to about 200 nM. The concentration depends upon the individual compound and the state of the neoplastic cells.

35 The method may also comprise initially treating the cells with an antitumor agent so as to render them resistant to an antitumor agent and subsequently contacting the resulting

resistant cells under suitable conditions with an effective amount of any of the compounds above, effective to selectively induce terminal differentiation of such cells.

5 The present invention also provides a method of treating a patient having a tumor characterized by proliferation of neoplastic cells which comprises administering to the patient an effective amount of any of the compounds above, effective to selectively induce growth arrest, terminal differentiation 10 and/or apoptosis of such neoplastic cells and thereby inhibit their proliferation.

The method of the present invention is intended for the treatment of human patients with tumors. However, it is also 15 likely that the method would be effective in the treatment of tumors in other mammals. The term tumor is intended to include any cancer caused by the proliferation of neoplastic cells, such as prostate cancer, lung cancer, acute leukemia, multiple myeloma, bladder carcinoma, renal carcinoma, breast carcinoma, 20 colorectal carcinoma, neuroblastoma or melanoma.

Routes of administration for the compound of the present invention include any conventional and physiologically acceptable route, such as, for example, oral, pulmonary, 25 parenteral (intramuscular, intraperitoneal, intravenous (IV) or subcutaneous injection), inhalation (via a fine powder formulation or a fine mist), transdermal, nasal, vaginal, rectal, or sublingual routes of administration and can be formulated in dosage forms appropriate for each route of 30 administration.

The present invention also provides a pharmaceutical composition comprising a pharmaceutically acceptable carrier, such as sterile pyrogen-free water, and a therapeutically acceptable 35 amount of any of the compounds above. Preferably, the effective amount is an amount effective to selectively induce terminal

-26-

differentiation of suitable neoplastic cells and less than an amount which causes toxicity in a patient.

The present invention provides the pharmaceutical composition 5 above in combination with an antitumor agent, a hormone, a steroid, or a retinoid.

The antitumor agent may be one of numerous chemotherapy agents such as an alkylating agent, an antimetabolite, a hormonal 10 agent, an antibiotic, colchicine, a vinca alkaloid, L-asparaginase, procarbazine, hydroxyurea, mitotane, nitrosoureas or an imidazole carboxamide. Suitable agents are those agents which promote depolarization of tubulin. Preferably the antitumor agent is colchicine or a vinca 15 alkaloid; especially preferred are vinblastine and vincristine.

In embodiments where the antitumor agent is vincristine, an amount is administered to render the cells are resistant to vincristine at a concentration of about 5 mg/ml. The administration of the agent is performed essentially as 20 described above for the administration of any of the compounds. Preferably, the administration of the agent is for a period of at least 3-5 days. The administration of any of the compounds above is performed as described previously.

25 The pharmaceutical composition may be administered daily in 2-6 hour infusions for a period of 3-21 days, for example, daily in a 4 hour infusion for a period of 5 days.

This invention will be better understood from the Experimental 30 Details which follow. However, one skilled in the art will readily appreciate that the specific methods and results discussed are merely illustrative of the invention as described more fully in the claims which follow thereafter.

EXPERIMENTAL DETAILS

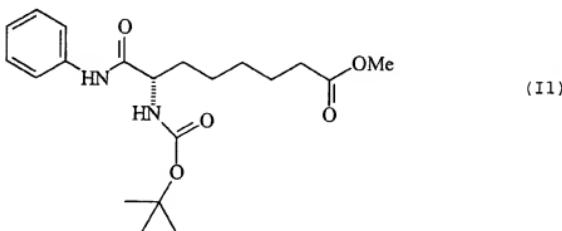
Examples 1-5 show the synthesis of substituted L- α -aminosuberic hydroxamic acids according to the subject invention, and 5 Examples 6 and 7 show the effects of compounds 1-5 on MEL cell differentiation and Histone Deacetylase activity.

Example 1 - Synthesis of Compound 1

10 **N-Boc- ω -methyl-(L)- α -aminosuberate, Boc-Asu(OMe)** was prepared according to a published procedure (40). ("Boc" = t-butoxycarbonyl; "Asu" = α -aminosuberate (or α -aminosuberic acid))

15 **N-Cbz- ω -t-butyl-(L)- α -aminosuberate, dicyclohexylamine salt** was purchased from Research Plus, Bayonne, NJ.

N-Boc- ω -methyl-(L)- α -aminosuberateanilide, Boc-Asu(OMe)-NHPH.



20

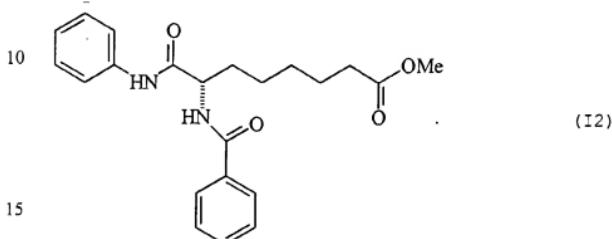
N-Boc- ω -methyl-(L)- α -aminosuberate (493mg, 1.63mmoles) was dissolved under Ar in 7mL of dry CH_2Cl_2 . EDC (470mg, 2.45mmoles) was added, followed by aniline (230 μ L, 2.52 mmoles). The solution was stirred at room temperature for 2h 30min, then 25 washed with dilute HCl (pH 2.4, 2x5mL), sat. NaHCO_3 (10mL), and H_2O (2x10mL). The product was purified by column chromatography

(Silica gel, Hexanes: AcOEt 3.5:1). The isolated yield was 366mg (60%).

¹H-NMR and Mass Spectroscopy were consistent with the product.

5

N-Benzoyl- ω -methyl-(L)- α -aminosuberanilide, PhCOHN-Asu(OMe)-NHPh.

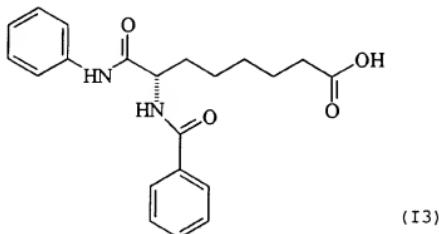


20 90mg of N-Bloc- ω -methyl-(L)- α -aminosuberanilide (0.238mmoles) were treated with 3.2mL of 25% trifluoroacetic acid (TFA) CH₂Cl₂ for 30 min. The solvent was removed and the residue left under high vacuum for 12h. It was dissolved under Ar in 3mL of dry CH₂Cl₂ and benzotriazole-1-yloxy-tris-pyrrolidinophosphonium 25 hexafluorophosphate (PyBOP) (149mg, 0.286mmoles), benzoic acid (44mg, 0.357mmoles) and diisopropylethylamine (114 μ L, 0.655mmoles). The solution was stirred at room temperature for 1h. The product was purified by column chromatography (Silica gel, Hexanes: AcOEt 3:1-2:1) as a white solid: 75mg, 82%.

30

¹H-NMR and Mass Spectroscopy were consistent with the product.

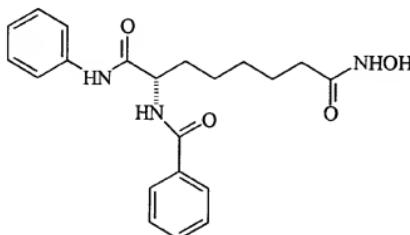
The foregoing coupling reaction was also successfully accomplished using 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide 35 hydrochloride (EDC) as a reagent.

N-Benzoyl-(L)- α -aminosuberoylanilide, PhCONH-Asu(OH)-NHPH.

5 75mg (0.196mmoles) of N-benzoyl--aminosuberateanilide were stirred for 6h at 0°C in 1M NaOH:THF:MeOH 1:1:1. After complete disappearance of the starting material, the solution was neutralized (1M HCl) and extracted with AcOEt. The organic phase was collected and dried. Solvent removal yielded the 10 product as a white solid: 67mg, 93%.

¹H-NMR and Mass Spectroscopy were consistent with the product.

**N-Benzoyl-(L)- α -aminosuberoylanilide- ω -hydroxamic acid,
15 PhCONH-Asu(NHOH)-NHPH:**



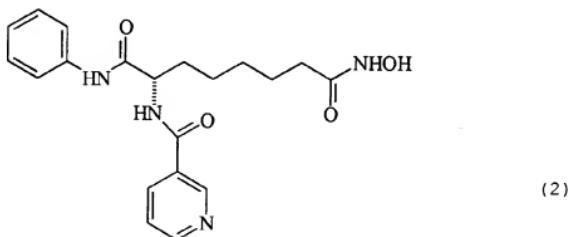
To a suspension of 26mg of N-benzoyl- ω -methyl-(L)- α -aminosuberanilide (I2) in 1mL of dry CH_2Cl_2 was added 58mg of $\text{H}_2\text{NOTBDPS}$ ($\text{H}_2\text{NO-t-butylidiphenylsilyl}$) followed by 22mg of EDC. The reaction was stirred at room temperature for 4h. The intermediate protected hydroxamic acid was purified by column chromatography (silica gel, CH_2Cl_2 : MeOH 100:0-98-2). It was deprotected by treatment with 5% TFA in CH_2Cl_2 for 1h30min. The product was precipitated from acetone-pentane.

25 $^1\text{H-NMR}$ (d_6 -DMSO, 500MHz) δ = 10.29 (s, 1H), 8.53 (d, 1H), 7.90 (d, 2H), 7.60 (d, 2H), 7.53 (m, 1H), 7.46 (t, 2H), 7.28 (t, 2H), 7.03 (t, 2H), 4.53 (q, 1H), 1.92 (t, 2H), 1.78 (m, 2H), 1.50-1.25 (m, 6H).

30 ESI-MS : 384 (M+1), 406 (M+Na), 422 (M+K)

Example 2 - Synthesis of Compound 2

N-Nicotinoyl-(L)- α -aminosuberoylanilide- ω -hydroxamic acid,
35 $\text{C}_5\text{H}_4\text{NCO-Asu(NHOH)-NHPh}$:



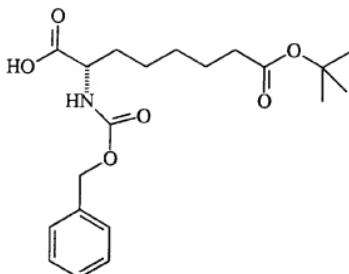
It was prepared from N-Boc- ω -methyl-L- α -aminosuberate following the same procedure used for the benzoyl analog. Yields and 40 chromatographic behaviour were comparable.

-31-

¹H-NMR (d₆-DMSO, 500MHz) δ= 10.30 (s, 1H), 10.10 (s, 1H), 9.05 (m, 1H), 8.80 (m, 1H), 8.71 (m, 1H), 8.24 (m, 1H), 7.60 (m, 2H), 7.30 (m, 2H), 7.04 (m, 1H), 4.56 (m, 1H), 1.93 (t, 2H), 1.79 (m, 2H), 1.55-1.30 (m, 6H). ESI-MS : 385 (M+1), 407 (M+Na).

Example 3 - Synthesis of Compound 3

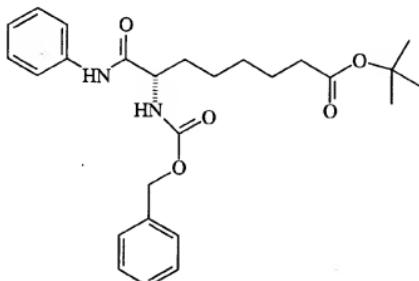
N-benzyloxycarbonyl- ω -t-butyl-(L)-amino suberic acid,
N-Cbz-(L)-Asu(OtBu)-OH.



(T4)

5 *N*-Cbz-(L)-Asu(OtBu)-OH, dicyclohexylamine salt (100 mg, 0.178 mmol) was partitioned between 1 M HCl (5mL) and EtOAc (10mL). The organic layer was removed, and the aqueous portion washed with EtOAc (3 x 3 mL). The organic fractions were combined, washed with brine (1 x 2 mL), and dried (MgSO_4). The mixture 10 was filtered and concentrated to a colorless film (67 mg, 0.176 mmol, 99%). This compound was used immediately in the next step.

15 N-Cbz-(L)-Asu(OtBu)-NHPh.

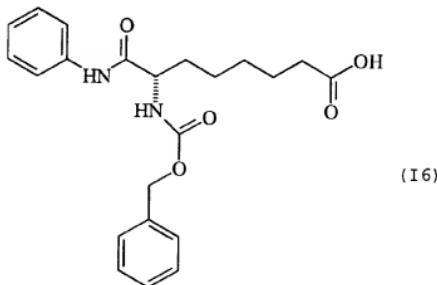


(15)

N-Cbz-(L)-Asu(OtBu)-OH (67mg, 0.176 mmol) was dissolved in dry CH₂Cl₂ (2.5 mL). Aniline (17 μ L, 0.187 mmol), PyBOP (97 mg, 0.187 mmol), and *i*Pr₂NEt (46 μ L, 0.266 mmol) were added and the mixture stirred for 2 h. The reaction was complete as indicated by TLC. The mixture was diluted with EtOAc (5 mL) and water (5 mL), and the layers separated. The aqueous portion was washed with EtOAc (3 x 3 mL) and the organic fractions combined. This solution was washed with 1 M HCl (1 x 2 mL) and brine (1 x 2 mL), dried (MgSO₄), filtered, and concentrated to a crude oil. This was passed through a plug of silica gel (30% EtOAc/hexanes) to remove baseline impurities, affording the compound (76mg, 0.167 mmol, 94%).

30 ¹H NMR (CDCl₃, 400 MHz, no TMS) δ 8.20 (br s, 1H), 7.47 (d, 2H), 7.32 (m, 5H), 7.28 (t, 2H), 7.08 (t, 1H), 5.39 (d, 1H), 5.10 (m, 2H), 4.26 (m, 1H), 2.18 (t, 2H), 1.93 (m, 1H), 1.67 (m, 1H), 1.55 (m, 3H), 1.42 (s, 9H), 1.36 (m, 3H).

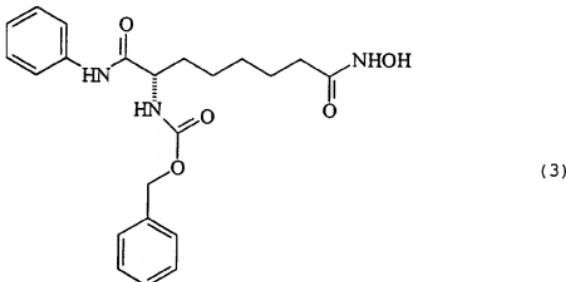
35 **N**-benzyloxycarbonyl-(L)- α -aminosuberateanilide,
N-Cbz-(L)-Asu(OH)-NHPhe.



N-Cbz-(L)-Asu(OtBu)-anilide (76mg, 0.167 mmol) was dissolved in dry CH_2Cl_2 (5 mL) and TFA (0.5 mL) added dropwise. The reaction was complete by TLC after 3h. The mixture was concentrated in vacuo to give the title compound (80 mg, crude). This compound 5 was taken on without purification to the next step.

^1H NMR (DMSO- d_6 , 400 MHz) δ 11.93 (br s, 1H), 9.99 (br s, 1H), 7.57 (m, 3H), 7.34 (m, 5H), 7.29 (t, 2H), 7.03 (t, 1H), 5.02 (m, 2H), 4.11 (m, 1H), 2.17 (t, 2H), 1.61 (m, 2H), 1.46 (m, 2H), 10 1.27 (m, 4H).

N-benzyloxycarbonyl-(L)- α -aminosuberateanilide ω -hydroxamic acid, N-Cbz-(L)-Asu(NH-OH)-NHPH.



15

N-Cbz-(L)-Asu(OH)-anilide (80 mg, crude) and O-*t*-butyldiphenylsilyl-hydroxylamine (60 mg, 0.221 mmol) were dissolved in CH_2Cl_2 (4 mL). To this was added PyBOP (125 mg, 0.241 mmol) and *iPr*₂NET (52 μL , 0.302 mmol) and stirred 20 overnight. TLC indicated reaction completion. The mixture was concentrated in vacuo and then passed through a plug of silica gel (50% EtOAc/hexanes) to remove baseline impurities. Evaporation of volatiles afforded 107 mg of material which was then dissolved in dry CH_2Cl_2 (5mL) and TFA (0.25 mL) was added. 25 Monitoring by TLC indicated completion after 1.5h. Concentrated

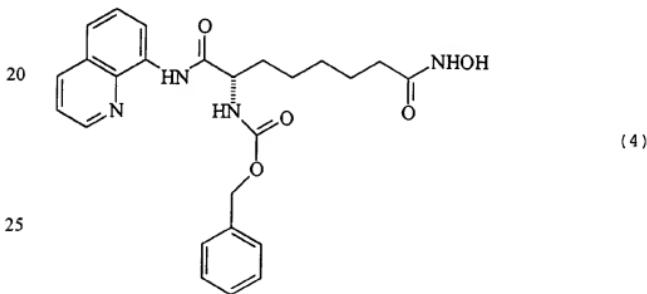
-35-

in *vacuo* to remove all volatiles. The residue was taken up in EtOAc (3mL), and then hexanes was added slowly to result in the precipitation of a white gel. The supernatant was removed, and the precipitate washed with hexanes (3 x 2 mL). This material was taken to dryness under reduced pressure, to afford the title compound (40 mg, 0.097 mmol, 59%).

¹H NMR (DMSO-d₆, 400 MHz) δ 10.32 (s, 1H), 10.00 (s, 1H), 8.64 (br s, 1H), 7.57 (m, 3H), 7.37 (m, 5H), 7.30 (t, 2H), 7.04 (t, 10 1H), 5.02 (m, 2H), 4.12 (m, 1H), 1.93 (t, 2H), 1.62 (m, 2H), 1.45 (m, 2H), 1.29 (m, 4H); ESI-MS 414 (M+1).

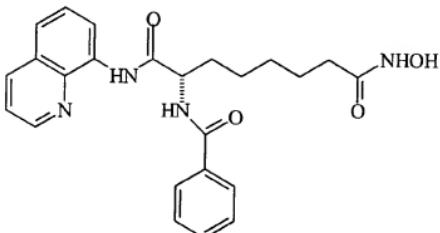
Example 4 - Synthesis of Compound 4

15 N-benzyloxycarbonyl-(L)- α -aminooxoberoyl-8-quinolinamide- ω -hydroxamic acid.



30 Prepared in similar manner to compound 3.

1H NMR (DMSO-d₆, 400 MHz) δ 10.45 (s, 1H), 10.31 (s, 1H), 8.85 (dd, 1H), 8.63 (dd, 1H), 8.42 (dd, 1H), 8.13 (dd, 1H), 8.68 (m, 2H), 7.60 (t, 1H), 7.37 (m, 2H), 7.28 (m, 2H), 5.10 (m, 2H), 35 4.24 (m, 1H), 1.93 (t, 2H), 1.85 (m, 1H), 1.70 (m, 1H), 1.50 (m, 2H), 1.42 (m, 2H), 1.30 (m, 2H); ESI-MS 465 (M+1).

Example 5 - Synthesis of Compound 5**N-Benzoyl-(L)- α -aminosuberoyl-8-quinolinamide- ω -hydroxamic acid:**

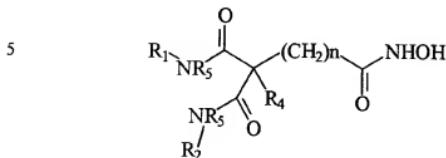
5

A sample of the N-Cbz- ω -t-butyl L- α -aminosuberoyl-8-quinolinamide (90mg, 0.178 mmoles) was obtained from the previous synthesis. The Cbz group was removed by hydrogenation in MeOH on 5%Pd on C. The resulting free amine was coupled with benzoic acid using EDC in dry CH_2Cl_2 , (69% over the two steps). After TFA deprotection of the t-butyl ester, the usual coupling with $\text{H}_2\text{NOTBDPS}$ followed by deprotection afforded the desired hydroxamic acid.

$^1\text{H-NMR}$ (d_6 -DMSO, 500MHz) δ =10.55 (s, 1H), 10.30 (s, 1H), 9.03 (m, 1H), 8.78 (m, 1H), 8.62 (m, 1H), 8.40 (m, 1H), 7.97 (m, 2H), 7.67-7.46 (m, 6H), 4.66 (m, 1H), 1.94 (t, 2H), 1.87 (m, 1H), 1.80-1.20 (m, 7H). ESI-MS : 435 (M+1).

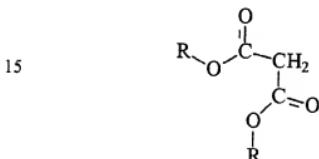
-37-

Example 6 - Synthesis of compound with inverted amide group.
 A compound having the following formula:

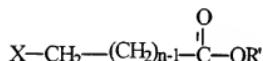


10

is synthesized by treating a malonic ester:

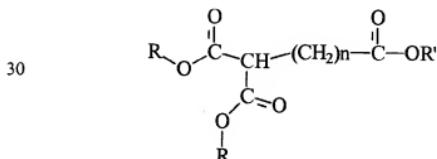


20 with a base, and then adding:

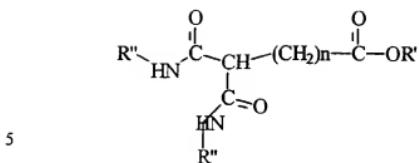


25

where X is a halogen, to form:



35 from which R is removed by reaction with an amine and a carbodiimide reagent to form:



from which R' is removed and converted to hydroxamic acid (NHOH) 10 as in the previous examples.

In the foregoing scheme, R may be t-butyl, removed with trifluoroacetic acid; R' may be methyl, removed with a base or LiI; and each R'' may be the same or different, depending on the 15 reagent used.

Example 7 - Effect of Compound 1 (N-Benzoyl-(L)- α -aminosuberoylanilide- ω -hydroxamic acid, PhCONH-Asu(NHOH)-NHPH) on MEL Cell Differentiation and Histone Deacetylase Activity

Murine erythroleukemia (MEL) cell differentiation.

The MEL cell differentiation assay was used to assess the ability of Compound 1 to induce terminal differentiation. MEL cells (logarithmically dividing) were cultured with the indicated concentrations of Compound 1. Following a 5-day culture period, cell growth was determined using a Coulter Counter and differentiation was determined microscopically using the benzidine assay to determine hemoglobin protein accumulation on a per cell basis.

30

It was observed, as shown in Figure 1, that Compound 1 (200nM) is able to induce MEL cell differentiation.

Histone Deacetylase (HDAC) enzymatic activity.

35 The effect of Compound 1 on affinity purified human epitope-tagged (Flag) HDAC1 was assayed by incubating the enzyme preparation in the absence of substrate on ice for 20 min with

the indicated amounts of Compound 1. Substrate ([³H]acetyl-labeled murine erythroleukemia cell-derived histone) was added and the samples were incubated for 20 min at 37°C in a total volume of 30 μ l. The reactions were then stopped and released 5 acetate was extracted and the amount of radioactivity released determined by scintillation counting.

It was observed, as shown in Figure 2, that Compound 1 is a potent inhibitor of HDAC1 enzymatic activity (ID_{50} =lnM).

10

Example 8 - Effect of Compound 2 (N-Nicotinoyl-(L)- α -aminosuberoylanilide- ω -hydroxamic acid, C_5H_4NCO -Asu(NHOH)-NHPH) on MEL Cell Differentiation

15 Murine erythroleukemia (MEL) cell differentiation:

The MEL cell differentiation assay was used to assess the ability of Compound 2 to induce terminal differentiation. MEL cells (logarithmically dividing) were cultured with the indicated concentrations of Compound 2. Following a 5-day 20 culture period differentiation was determined microscopically using the benzidine assay to determine hemoglobin protein accumulation on a per cell basis.

It was observed, as shown in Figure 3, that Compound 2 (800nM) 25 is able to induce MEL cell differentiation.

Example 9 - Effect of Compound 3 (N-benzyloxycarbonyl-(L)- α -aminosuberateanilide ω -hydroxamic acid, N-Cbz-(L)-Asu(NH-OH)-NHPH) on MEL Cell Differentiation and Histone Deacetylase 30 Activity

Murine erythroleukemia (MEL) cell differentiation:

The MEL cell differentiation assay was used to assess the ability of Compound 3 to induce terminal differentiation. MEL 35 cells (logarithmically dividing) were cultured with the indicated concentrations of Compound 3. Following a 5-day culture period differentiation was determined microscopically using the benzidine assay to determine hemoglobin protein accumulation on a per cell basis.

It was observed, as shown in Figure 4, that Compound 3 (400nM) is able to induce MEL cell differentiation.

Histone deacetylase (HDAC) enzymatic activity:

5 The effect of Compound 3 on affinity purified human epitope-tagged (Flag) HDAC1 was assayed by incubating the enzyme preparation in the absence of substrate on ice for 20 min with the indicated amounts of HPC. Substrate ($[^3\text{H}]$ acetyl-labelled murine erythroleukemia cell-derived histone) was added and the 10 samples were incubated for 20 min at 37°C in a total volume of 30 μl . The reactions were then stopped and released acetate was extracted and the amount of radioactivity released determined by scintillation counting.

15 It was observed, as shown in Figure 5, that Compound 3 is a potent inhibitor of HDAC1 enzymatic activity (ID_{50} =100 nM).

Example 10 - Effect of Compound 4 (N-benzyloxycarbonyl-(L)- α -aminooxoberoyl-8-quinolinamide- ω -hydroxamic acid) on MEL Cell Differentiation and Histone Deacetylase Activity

Murine erythroleukemia (MEL) cell differentiation:

The MEL cell differentiation assay was used to assess the ability of Compound 4 to induce terminal differentiation. MEL 25 cells (logarithmically dividing) were cultured with the indicated concentrations of Compound 4. Following a 5-day culture period differentiation was determined microscopically using the benzidine assay to determine hemoglobin protein accumulation on a per cell basis.

30

It was observed, as shown in Figure 6, that Compound 4 (40 nM) is able to induce MEL cell differentiation.

Histone deacetylase (HDAC) enzymatic activity:

35 The effect of Compound 4 on affinity purified human epitope-tagged (Flag) HDAC1 was assayed by incubating the enzyme preparation in the absence of substrate on ice for 20 min with indicated amounts of HPC. Substrate ($[^3\text{H}]$ acetyl-labelled murine

erythroleukemia cell-derived histone) was added and the samples were incubated for 20 min at 37°C in a total volume of 30 μ l. The reactions were then stopped and released acetate was extracted and the amount of radioactivity released determined by scintillation counting.

It was observed, as shown in Figure 7, that Compound 4 is a potent inhibitor of HDAC1 enzymatic activity ($ID_{50} < 10$ nM).

10 SAHA inhibits the activity of affinity purified HDAC1 and HDAC3 (39). Crystallographic studies with SAHA and a HDAC related protein reveal that SAHA inhibits HDAC by a direct interaction with the catalytic site (66). Additional studies demonstrate that a tritium labeled photoaffinity SAHA analog (3 H-498) that 15 contains an azide moiety (67) binds directly to HDAC1 (Fig. 8). These results indicate that this class of hydroxamic acid based compound inhibits HDAC activity through a direct interaction with the HDAC protein.

20 SAHA causes the accumulation of acetylated histones H3 and H4 in vivo. The in vivo effect of SAHA has been studied using the CWR22 human prostate xenograft in mice (68). SAHA (50 mg/kg/day) caused a 97% reduction in mean final tumor volume compared to controls with no apparent toxicity. SAHA 25 administration at this dose caused an increase in acetylated histones H3 and H4 in the tumor xenograft (Fig 9).

SAHA is currently in Phase I Clinical Trials in patients with solid tumors. SAHA causes an accumulation of acetylated 30 histones H3 and H4 in the peripheral blood mononuclear cells isolated from patients undergoing treatment (Fig. 10).

Table 1 shows a summary of the results of the Examples 7-10, testing compounds 1-4, and also compares the results to the results obtained from using SAHA.

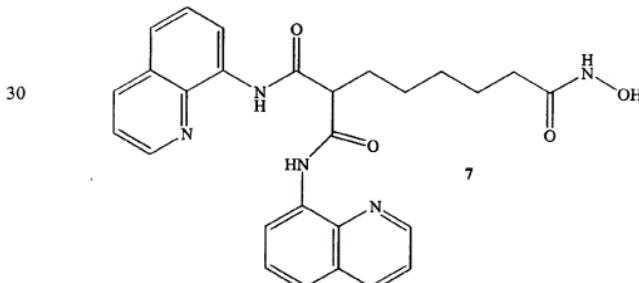
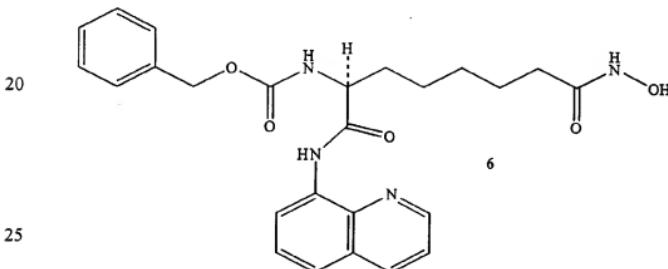
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Table 1. Summary of Test results of compounds 1-4, and comparison to SAHA results.

10	Compound	MEL Differentiation			HDAC Inhibition	
		Range	Opt.	%B+	Range	ID50
15	1	0.1 to 50 μ M	200 nM	44%	0.0001 to 100 μ M	1 nM
20	2	0.2 to 12.5 μ M	800 nM	27%		TBT
25	3	0.1 to 50 μ M	400 nM	16%	0.01 to 100 μ M	100 nM
30	4	0.01 to 50 μ M	40 nM	8%	0.01 to 100 μ M	<10 nM
	SAHA		2500 nM	68%	0.01 to 100 μ M	200 nM

Example 12 - Modified Inhibitors of HDAC

In additional studies we found that compounds **6** and **7** shown below were very effective inhibitors of the enzyme HDAC. Compound **6** had ID_{50} of 2.5 nM, and compound **7** had ID_{50} of 50 nM. This contrasts with an ID_{50} for SAHA of 1 μ M, much higher. Note that the 1 μ M ID_{50} for SAHA as an inhibitor of HDAC is of the same general magnitude as its 2.5 μ M optimal dose for the cytodifferentiation of MEL cells, but this close similarity is not true for all the compounds examined. In some cases very effective HDAC inhibitors are less effective as cytodifferentiators, probably because the drugs are metabolized in the cell assays. Also, all cell types are not the same, and some compounds are much better against human tumor cells such as HT-29 than they are against MEL cells. Thus, inhibition of HDAC cells is a preliminary indicator.



Example 13 - Evolution of Compounds without a Hydroxamic Acid Portion

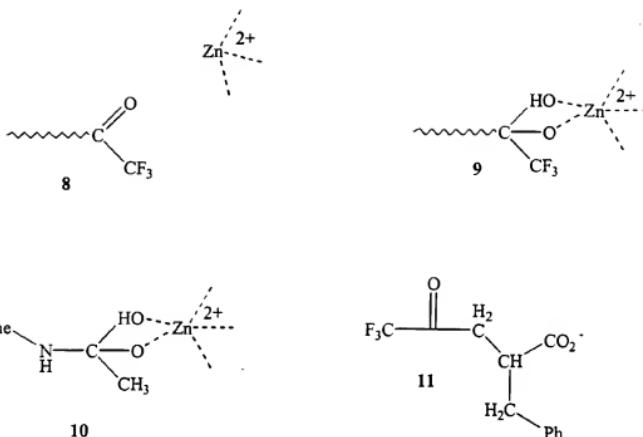
5 Of the above compounds which are hydroxamic acids, we have found that they undergo enzymatic hydrolysis rather rapidly to the carboxylic acids, so their biological lifetimes are short. We were interested in evolving compounds which might be more stable in vivo. Thus we have developed inhibitors of HDAC that are not 10 hydroxamic acids, and that can be used as cytodifferentiating agents with longer biological lifetimes. Furthermore, we found that the newly evolved compounds have better selectivity to HDAC than, e.g. SAHA.

15 We have evolved compounds that have double bonds, similarly to Trichostatin A (TSA) to see if the resulting compounds have even greater efficacy. Also, the chain in TSA is only five carbons, not the six of SAHA. In Oxamflatin there is a chain of four carbons containing a double bond and an ethinyl link between the 20 hydroxamic acid and the first phenyl ring, and Oxamflatin has been claimed to be an effective inhibitor of HDAC. We incorporate some of these features in our compounds, including those compounds that are not hydroxamic acids.

25 Also disclosed are simple combinatorial methods for screening a variety of such compounds for efficacy and selectivity with respect to HDAC inhibition.

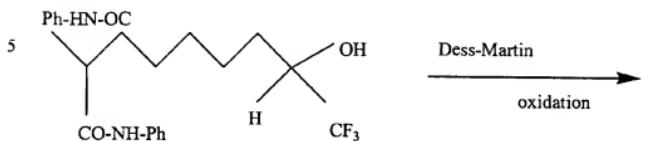
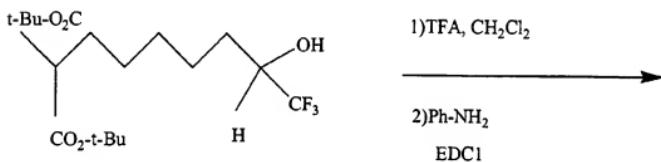
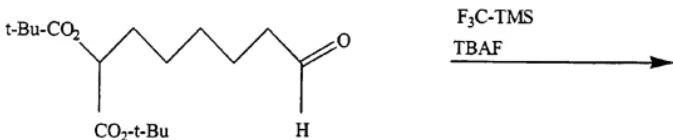
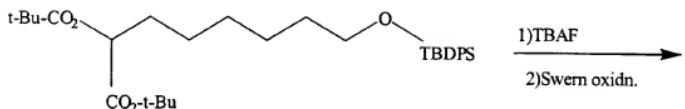
Furthermore, since there are many important enzymes that contain 30 Zn(II), hydroxamic acids, and perhaps some of the other metal coordinating groups, can also bind to Zn(II) and other metals.

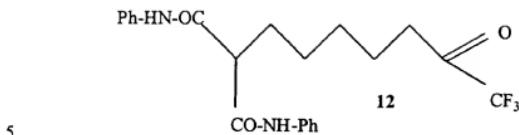
-45-



Since the target for HDAC is an acetyllysine sidechain of 5 histone, we make compounds in which transition state analogs of the substrate are present. For example, we synthesize compounds like SAHA in which the hydroxamic acid group $-\text{CO}-\text{NHOM}$ is replaced by a trifluoroacetyl group, $-\text{CO}-\text{CF}_3$. The resulting **8** will easily form a hydrate, and thus bind to the $\text{Zn}(\text{II})$ of HDAC **10** in a mimic **9** of the transition state **10** for deacetylation. This is related to the work published by Lipscomb [56] on the binding to carboxypeptidase A of a substrate analog **11** containing a $\text{CF}_3-\text{CO}-\text{CH}_2$ group in place of the normal amide. The hydrate of the ketone coordinated to the $\text{Zn}(\text{II})$ as a mimic of the transition state for catalyzed hydrolysis of an amide substrate. Our synthesis of a particular example **12** in the fluoroketone series is shown in Scheme below:

-46-





After the malonic ester alkylation, the aldehyde is prepared and then converted to the trifluoromethyl carbinol with Rupperts 10 reagent [57, 58]. The malonic bis-anilides are prepared, and the carbinol oxidized to the ketone 12 with the Dess-Martin reagent [59]. Other approaches were tried unsuccessfully. In particular, attempts to convert a carboxylic acid derivative directly to a trifluoromethyl ketone did not work.

15

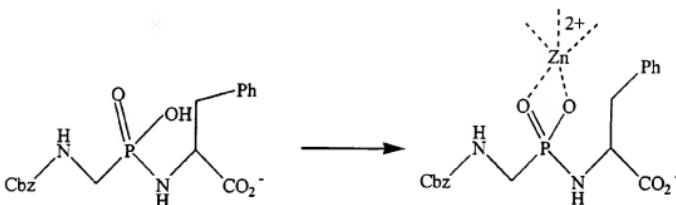
Compound 12 has been tested with HDAC and found to be an inhibitor of the enzyme. Thus, we also adapt this synthesis to the preparation of analogs of 12 with unsaturation, etc., in the chain, and other groups at the left end of the molecule.

20

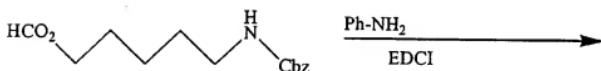
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Example 14 - Evolution of Compounds where the Hydroxamic Acid Group is Replaced by NH-P(O)OH-CH₃

An analog of SAHA in which the CH₂-CO-NHOH group is replaced by NH-P(O)OH-CH₃ may be synthesized by the general scheme shown 5 below. The resulting compound, **13**, binds to the Zn(II) of HDAC the way a related group binds to the Zn(II) of carboxypeptidase in analogs such as that prepared by Bartlett [60].



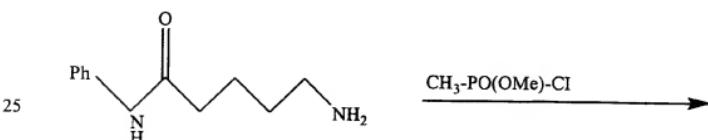
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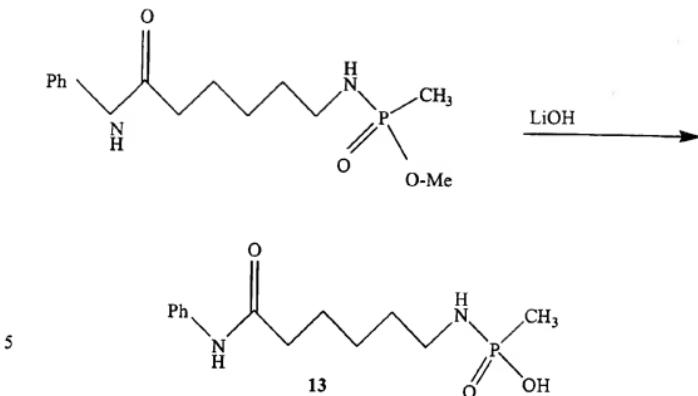


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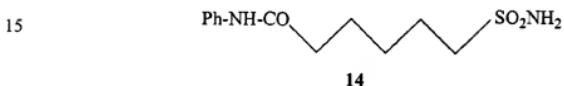
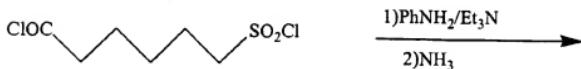
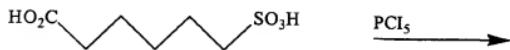
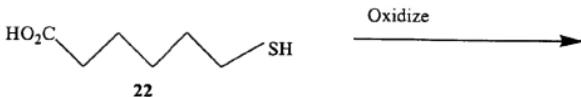
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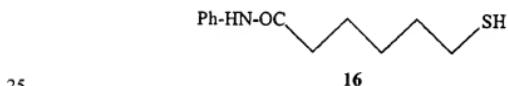


- 10 A classic inhibitor of the Zn(II) enzyme carbonic anhydrase is a sulfonamide, whose anion binds to the Zn(II) [61]. Thus compound **14**, an analog of SAHA with a sulfonamide group, is synthesized as shown below. In the last step we react a carboxylic sulfonic bis-chloride with aniline and ammonia.
- 15 Since the carboxylic acid chloride reacts faster, we use the sequence of aniline, then ammonia, but the sequence may be reversed, or the mixture may be separated if the two are of similar reactivity.
- 20 In the course of the synthesis of **14**, we use a thiol **15** easily made from the corresponding haloacid. Thiols are also inhibitors of Zn(II) enzymes such as carboxypeptidase A and related peptidases such as Angiotensin Converting Enzyme (ACE), so we convert **15** to **16** as an inhibitor of HDAC. A similar 25 synthesis can be used to attach the NH-P(O)OH-CH₃ group to other compounds, in particular compounds **6** and **7**.

-50-



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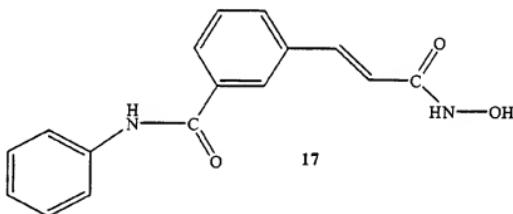


Example 15 - Varying the linker between the Zn(II) binding group and the hydrophobic binding groups.

Based on the results with Oxamflatin, it seems that a phenyl ring can be part of the chain between the Zn(II) binding group and the left hand section of the molecule as drawn, particularly when the phenyl ring is meta substituted. Thus, we provide a synthesis to incorporate such meta substituted chains into other of our compounds. We construct compounds 17 and 18. The simple syntheses, not shown in detail, only require that instead of the hydroxamic acid attached to the phenyl ring we make the aryl amides of 17 and 18.

15

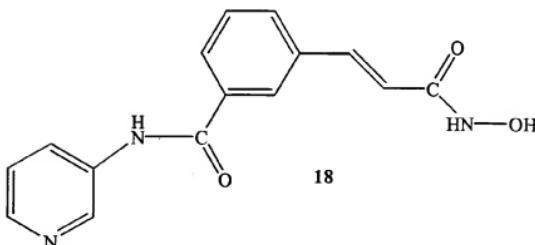
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Additional compounds may be synthesized, such as 19 and 20 to incorporate the trifluoromethyl ketone group of 12 that we know is effective as a Zn(II) binder in HDAC. The syntheses involve preparing compounds 21 and 22 and then adding CF₃ to form the

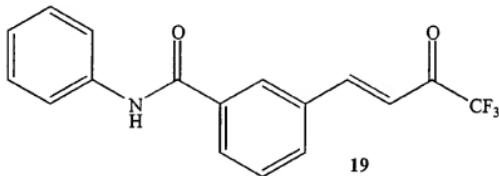
carbinol, followed by oxidation as in the synthesis of **12**. A simple synthesis involves Heck coupling of compounds **23** and **24** with ethyl acrylate, and conversion of the ester to aldehydes **21** and **22** by reduction to the carbinol and then reoxidation.

5

All the chains shown so far contain only carbon atoms, but thioether links may be acceptable and even useful, and they add synthetic ease. Thus, sulfonamides such as **25** and **26**, related to **19** and **20**, from the corresponding thiophenol and **10** bromomethylsulfonamide. A related synthesis may be used to make the corresponding phosphonamides **27** and **28**, if this class proves to be useful HDAC inhibitors and cytodifferentiators. In this case, (N-protected) m-aminobenzoic acid is used to acylate the arylamines, then phosphorylate the anilino group.

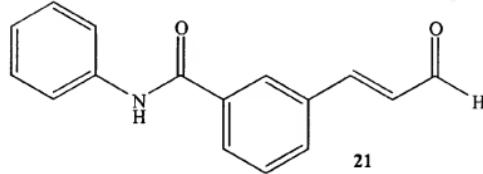
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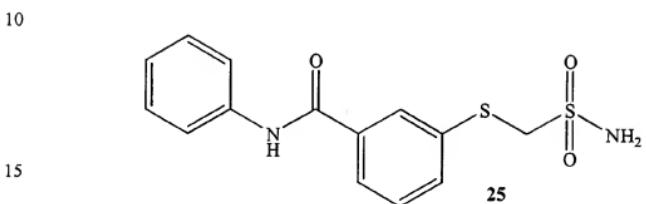
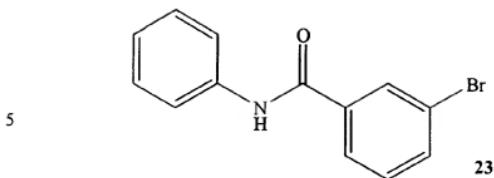


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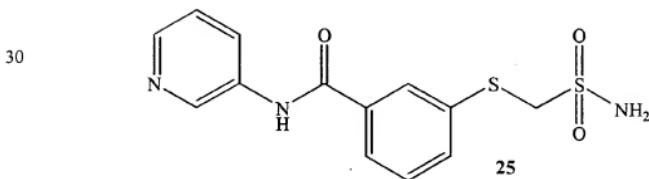
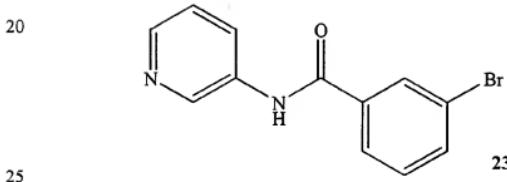
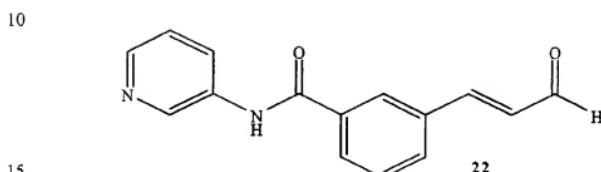
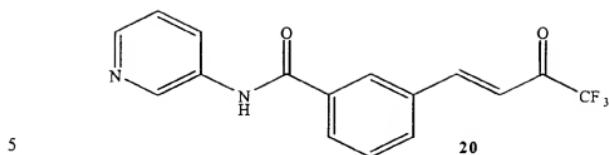


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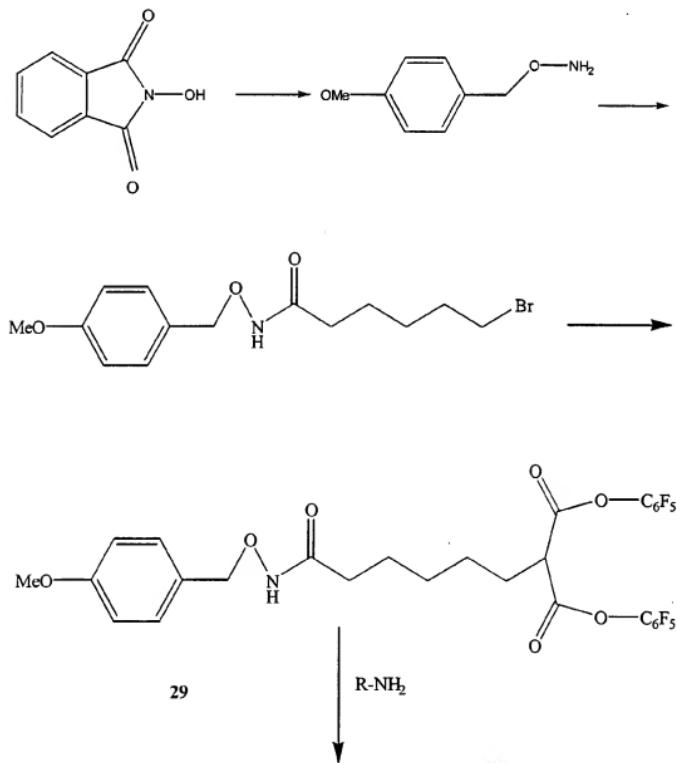
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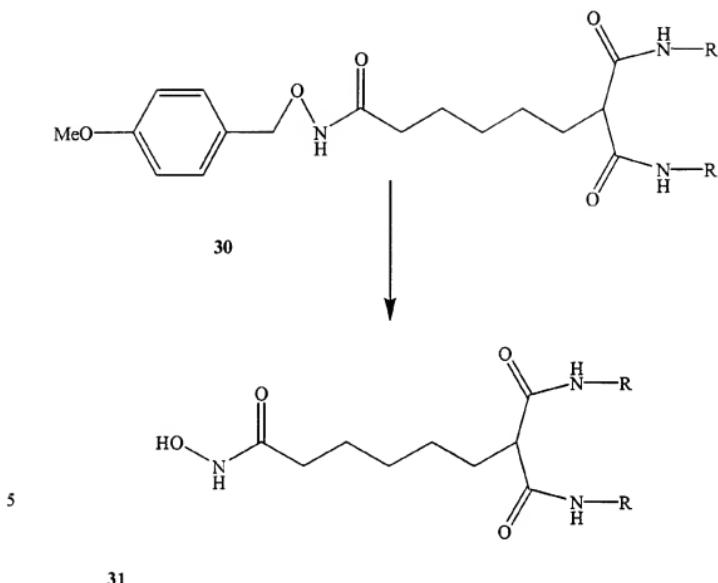
-54-



Example 16 - Varying the left hand of the molecule, carrying the hydrophobic groups.

To vary the hydrophobic groups, we synthesized compound **29**, as an intermediate that can be treated with various amines to make the compounds **30**. Then deprotection of the hydroxamic acid group will generate the general class **31**. The synthesis is shown in the scheme below.



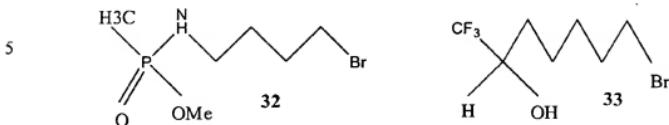


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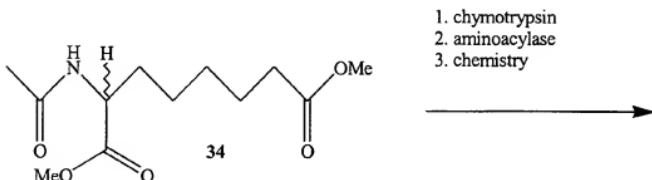
In the synthesis the O-protected hydroxylamine is acylated with bromohexanoic acid, and the compound then alkylates the bis-15 pentafluoro ester of malonic acid. The resulting **29** then reacts with various amines, and the protecting group is removed with acid.

With this compound as the starting material, we synthesize 20 related libraries carrying the other Zn(II) binding groups. For example, alkylation of the malonate with compound **32** lets us make a phosphonamidate library, and compound **33** will let us make a $\text{CF}_3\text{-CO}$ library. In a similar way, a sulfonamide library can be made if the work described earlier indicates that this is a 25 promising Zn(II) binding group for HDAC. Of course after

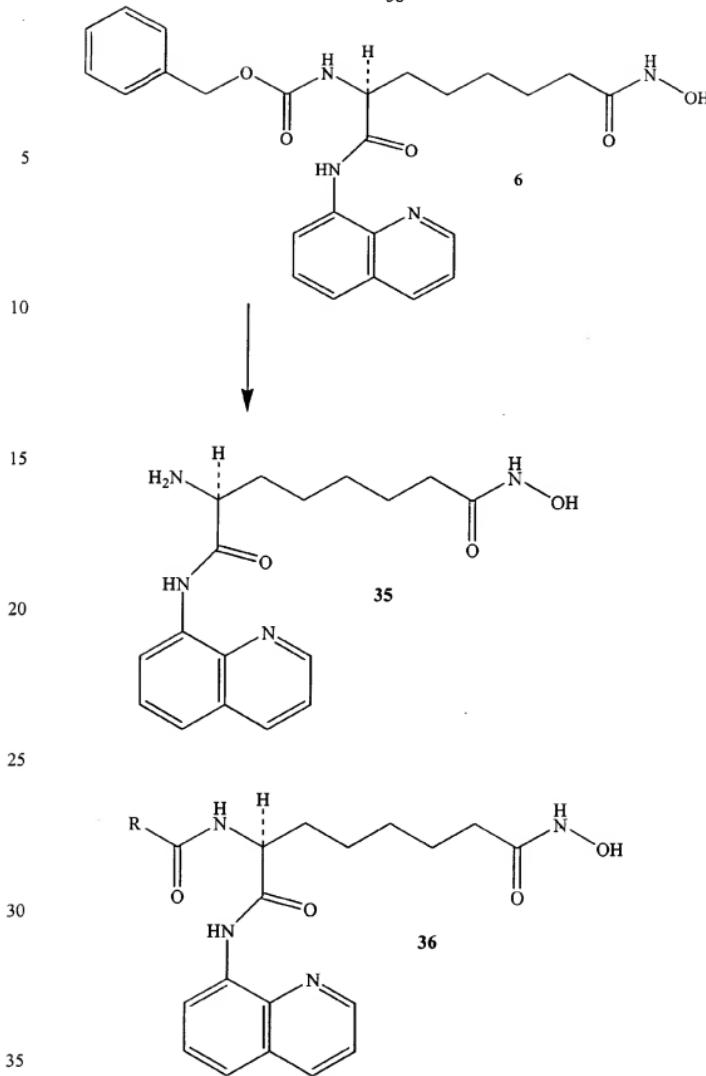
malonate alkylation and aminolysis the compound from **32** will be demethylated, while that from **33** will be oxidized.



10 This also allows to expand on the structure of compound **6**, the derivative of aminosuberic acid. As described, this was one of the most effective HDAC inhibitor we have examined. We prepared this compound using an enzymatic hydrolysis to achieve optical resolution and selectivity among the two carbomethoxy groups of 15 **34**, so that we could convert one of them to the aminoquinoline amide of **6** while protecting the nitrogen as a carbobenzoxy group. At the end of the synthesis we converted the remote carbomethoxy group to a hydroxamate. However, **6** is an intermediate that can be used to prepare other derivatives. The 20 carbobenzoxy group from **6** can be removed and the amine **35** can be acetylated with a variety of carboxylic acids to prepare library **36**, or sulfonic acid chlorides to prepare the corresponding sulfonamides.

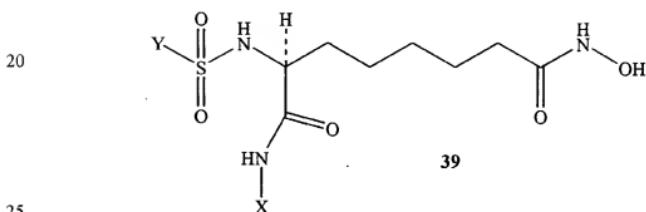
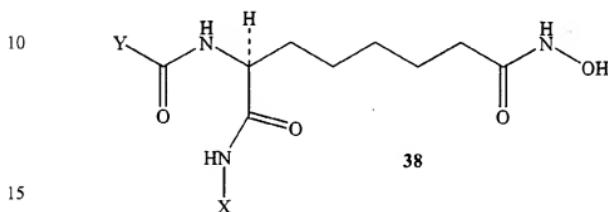
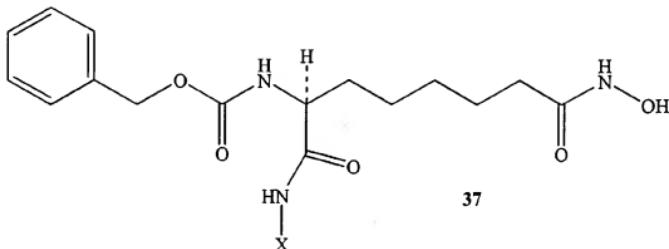


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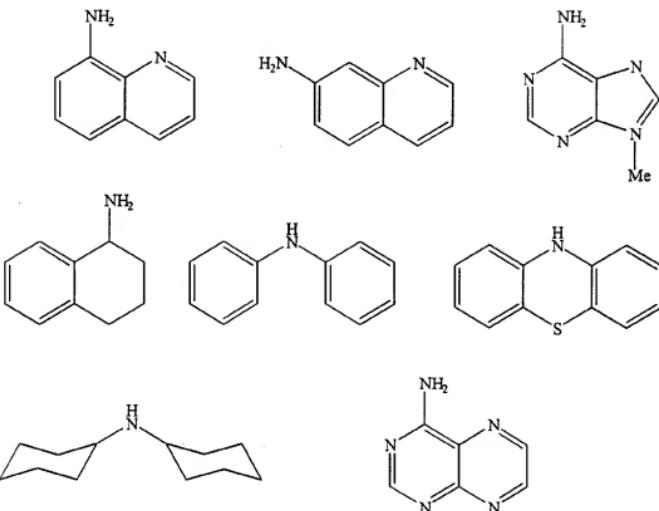
-59-

Also, we synthesize a different library of amides **37** related to **6**, and then expand it with a library of other amides **38** by acylating the amino group after deprotection. We also synthesize a group of compounds **39** in which after the 5 carbobenzoxy group of **37** is removed we make a library of sulfonamides using various sulfonyl chlorides. In all this, it the hydroxamic acid group may be protected.

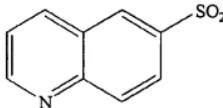
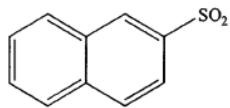
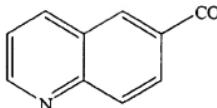
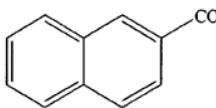
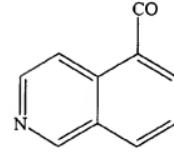
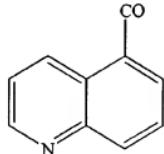
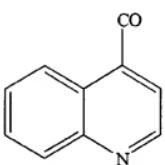
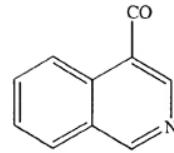
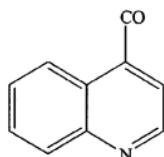
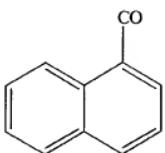


The foregoing synthesis schemes can be used to generate compounds having a large number of variation. Some substituent groups that are likely to result in compounds having potential good affinity to HDAC or having got differentiating activity are 5 as follows:

Some Amines that can be incorporated in place of the aniline in SAHA, or as the X group in compounds 37 and 38:



Some carboxylic and sulfonic acids that can be incorporated as group Y-CO in compound 38 or 39:



Example 17 - Synthesis using the foregoing schemes.

Reagents and starting materials were obtained from commercial suppliers and used without further purification unless otherwise indicated. For moisture-sensitive reactions, solvents were freshly distilled prior to use: tetrahydrofuran was distilled under argon from sodium metal utilizing benzophenone as an indicator; dichloromethane and acetonitrile were distilled from powdered calcium hydride. Anhydrous benzene, anhydrous DIEA, 10 and anhydrous pyridine were drawn by syringe from a sealed bottle purchased from Aldrich. *tert*-Butanol was dried over 4A molecular sieves before use. Sodium hydride was purchased as a 60% dispersion in mineral oil. Aniline, diisopropylamine, *N*-methylaniline, and benzyl alcohol were freshly distilled before 15 use. Deuterated solvents were obtained from Cambridge Isotope Laboratories. Air- and/or moisture-sensitive reactions were carried out under an atmosphere of dry argon in oven- or flame-dried glassware equipped with a tightly-fitting rubber septum. Syringes and needles were oven-dried before use. Reactions at 20 0 °C were carried out in an ice/water bath. Reactions at -78 °C were carried out in a dry ice/acetone bath.

Chromatography

Analytical thin-layer chromatography (TLC) was conducted on 25 glass plates precoated with silica gel 60 F-254, 0.25 mm thickness, manufactured by EM Science, Germany. Eluted compounds were visualized by one or more of the following: short-wave ultraviolet light, I₂ vapor, KMnO₄ stain, or FeCl₃ stain. Preparative TLC was carried out on Whatman precoated 30 plates of either 500 μm or 1000 μm silica gel thickness. Flash column chromatography was performed on Merck Kieselgel 60, 230-400 mesh.

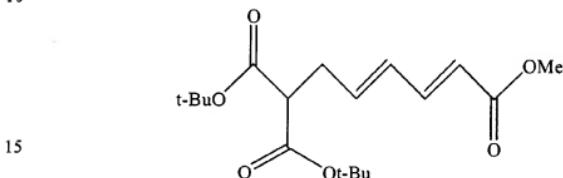
Instrumentation

35 NMR spectra were measured on Bruker DPX300 and DRX400 spectrometers; ¹H was observed at 300 and 400 MHz, and ¹⁹F at 376 MHz. Chemical shifts are reported as δ values in ppm relative

to the solvent residual peak. Mass spectra were obtained on a Nermag R-10-1 instrument for chemical ionization (CI) or electron impact ionization (EI) spectra, and on a Jeol JMS LCmate for electrospray ionization (ESI+) spectra. CI spectra were run with either ammonia (NH_3) or methane (CH_4) as the ionization gas.

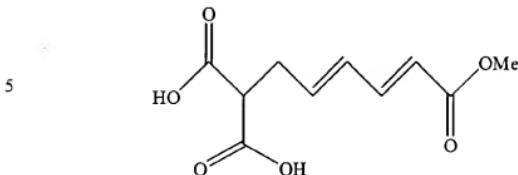
(E,E)-7-t-Butoxycarbonyl-octa-2,4-dienedioic acid 8-t-butyl ester 1-methyl ester (40)

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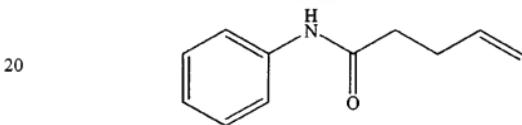
To a stirred solution of NaH (60% disp., 234 mg, 5.85 mmol) in THF (35 mL) at 0 °C was added di-*t*-butyl malonate (1.20 mL, 5.37 mmol) dropwise. Gas evolution was observed, and the solution was allowed to warm to ambient temperature and stirred for 6 h. A solution of methyl 6-bromo-2,4-hexadienoate (62) (1.00 g, 4.88 mmol) in THF (20 mL) was prepared in a separate flask and stirred in a water bath. To this was cannulated dropwise the 25 malonate mixture, and the reaction allowed to proceed overnight. The reaction was quenched with sat. NH_4Cl (5 mL), then H_2O (10 mL) was added and the mixture extracted with Et_2O (3 x 15 mL). The organic fractions were combined and washed with H_2O (1 x 10 mL), then with brine, dried over MgSO_4 , and filtered. 30 Evaporation under reduced pressure followed by flash chromatography (0-20% $\text{EtOAc}/\text{hexanes}$) gave **40** as a clear colorless oil (850 mg, 2.49 mmol, 51%). TLC R_f 0.66 (20% $\text{EtOAc}/\text{hexanes}$); $^1\text{H-NMR}$ (CDCl_3 , 400 MHz) δ 7.26 (dd, 1H), 6.26 (dd, 1H), 6.10 (m, 1H), 5.82 (d, 1H), 3.78 (s, 3H), 3.12 (t, 3H), 2.64 (t, 2H), 1.41 (s, 18H).

(E,E)-7-Carboxy-octa-2,4-dienedioic acid 1-methyl ester (41)



10 To a stirred solution of **40** (200 mg, 0.59 mmol) in CH_2Cl_2 (10 mL) was added TFA (1 mL). The reaction was allowed to proceed overnight. Volatiles were removed under reduced pressure to leave **41** as a white solid (112 mg, 0.49 mmol, 83%). $^1\text{H-NMR}$ (CD_3OD , 400 MHz) δ 7.11 (dd, 1H), 6.33 (dd, 1H), 6.16 (m, 1H), 15 5.81 (d, 1H), 3.76 (s, 3H), 3.15 (t, 1H), 2.70 (t, 2H).

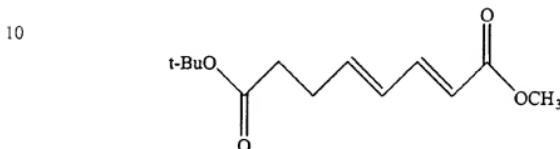
4-Pentenoic acid phenylamide (42)



To a stirred solution of oxalyl chloride (2.0 M in CH_2Cl_2 , 11.5 25 mL, 23.1 mmol) in CH_2Cl_2 (100 mL) and DMF (1 drop) at 0 °C was added 4-pentenoic acid (2.25 mL, 22.0 mmol). This was allowed to warm to ambient temperature. Upon cessation of gas evolution, the mixture was returned to 0 °C and a solution of aniline (2.00 mL, 22.0 mmol) and TEA (6.72 mL, 26.3 mmol) in 30 CH_2Cl_2 (5 mL) was added dropwise. After warming to ambient temperature, the reaction was allowed to proceed for 3 h. The mixture was concentrated under reduced pressure, and then partitioned between HCl (1 N, 10 mL) and EtOAc (30 mL) and the layers separated. The aqueous portion was extracted with EtOAc 35 (3 x 15 mL) and the organic layers combined, washed with brine, dried over MgSO_4 , and filtered. Concentration under reduced

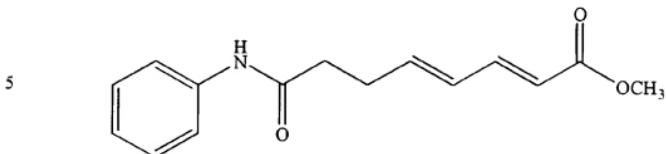
pressure gave a yellowish solid, which was recrystallized with toluene to obtain **42** as white crystals (1.97 g, 11.24 mmol, 51%). TLC R_f 0.68 (50% EtOAc/hexanes); $^1\text{H-NMR}$ (300 MHz, CDCl_3) δ 7.49 (d, 2H), 7.29 (t, 2H), 7.08 (t, 1H), 5.88 (m, 1H), 5.10 5 (dd, 2H), 4.42 (br s, 4H).

(E,E)-Octa-2,4-dienedioic acid 8-t-butyl ester 1-methyl ester (43)



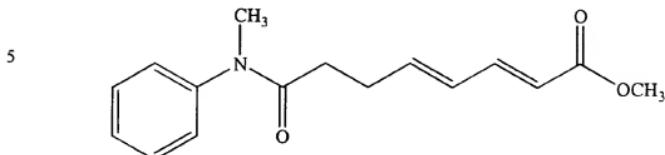
To a stirred solution of diisopropylamine (2.06 mL, 14.7 mmol) in THF (25 mL) at -78 °C was added *n*-BuLi (2.0 M in hexanes, 6.2 mL, 12.4 mmol) and allowed to stir 20 min at this temperature. A solution of phosphonate **43a** (63) (2.66 g, 11.3 mmol) in THF 20 (4 mL) was then added dropwise, giving a deep yellow color upon addition. After 20 min at -78 °C, the mixture was warmed to 0 °C and a solution of aldehyde **43b** (64) (1.78 g, 11.3 mmol) in THF (4 mL) was added dropwise. After addition the solution was allowed to warm to ambient temperature and stirred overnight. 25 It was diluted with Et_2O (30 mL) and washed with H_2O (3 x 10 mL). The aqueous washings were combined and extracted with Et_2O (2 x 10 mL), and the organic portions combined, washed with brine, dried over MgSO_4 , and filtered. Evaporation under reduced pressure followed by flash chromatography (10-20% EtOAc/hexanes) 30 gave **43** as a clear oil (1.54 g, 57%). TLC R_f 0.56 (20% EtOAc/hexanes); $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 7.22 (dd, 1H), 6.19 (dd, 1H), 6.08 (m, 1H), 5.77 (d, 1H), 2.42 (m, 2H), 2.32 (t, 2H), 1.42 (s, 9H).

(E,E)-7-Phenylcarbamoyl-hepta-2,4-dienoic acid methyl ester (44)



10 To a stirred solution of diester **43** (1.00 g, 4.61 mmol) in CH₂Cl₂ (40 mL) was added TFA (4.0 mL) and let react for 6 h. The mixture was concentrated under reduced pressure to remove volatiles. A white solid consisting of the crude acid (710 mg, 3.85 mmol) remained. This acid (400 mg, 2.17 mmol) was 15 dissolved in CH₂Cl₂ (20 mL) and to this stirred solution were added DMAP (13 mg), aniline (218 μ L, 2.39 mmol), and EDC (500 mg, 2.61 mmol). After 1.5 h, the mixture was diluted with EtOAc and washed with H₂O. The layers were separated, and the aqueous extracted with EtOAc (3 x 15 mL). The organic portions were 20 combined and washed with HCl (1 N, 1 x 5 mL) and brine, dried over MgSO₄, and filtered. Concentration under reduced pressure left a brown solid. This was dissolved in a minimum of CH₂Cl₂, then passed through a plug of silica gel (20-30% EtOAc/hexanes, 200 mL) to remove baseline impurities. The eluent was 25 concentrated to a light brown oil which was taken up in a small amount of CH₂Cl₂ and from which crystals were precipitated upon the addition of hexanes/diethyl ether. The mother liquor was drawn off, the crystals rinsed with ether, and the liquid fraction concentrated and this procedure repeated several times 30 to ultimately give **44** as off-white crystals (324 mg, 1.25 mmol, 58%). TLC R_f 0.44 (50% EtOAc/hexanes); ¹H-NMR (400 MHz, CDCl₃) δ 7.47 (d, 1H), 7.30 (t, 2H), 7.24 (m, 1H), 7.09 (t, 1H), 6.24 (dd, 1H), 6.14 (m, 1H), 5.81 (d, 1H), 3.72 (s, 3H), 2.60 (m, 2H), 2.47 (t, 2H).

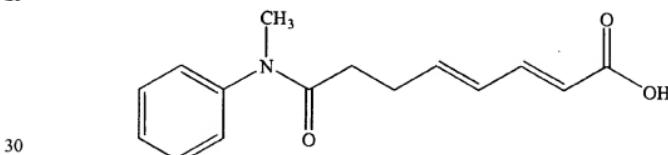
(E,E)-7-(Methyl-phenyl-carbamoyl)-hepta-2,4-dienoic acid methyl ester (45)



10 The crude acid intermediate from the first step of the preparation of **44** (200 mg, 1.09 mmol) and *N*-methylaniline (130 μ L, 1.19 mmol) were dissolved in CH_2Cl_2 (10 mL) and stirred. EDC (271 mg, 1.41 mmol) and DMAP (5 mg) were then added and the reaction run overnight. The mixture was partitioned between H_2O and EtOAc and the layers separated. The aqueous layer was extracted with EtOAc (3 x 10 mL), the organic portions combined and washed with HCl (1 N, 1 x 5 mL), then brine, dried over MgSO_4 , and filtered. Evaporation under reduced pressure left pure **45** as a brown oil (286 mg, 1.05 mmol, 96%). TLC R_f 0.81
 15 (5% MeOH/ CH_2Cl_2); $^1\text{H-NMR}$ (300 MHz, CDCl_3) δ 7.40 (t, 2H), 7.35 (t, 1H), 7.20 (d, 2H), 7.15 (dd, 1H), 6.20 (m, 2H), 5.76 (d, 1H), 3.70 (s, 3H), 3.24 (s, 3H), 2.42 (m, 2H), 2.18 (t, 2H).

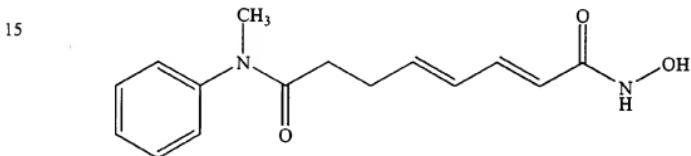
(E,E)-7-Phenylcarbamoyl-hepta-2,4-dienoic acid (46)

25



Ester **45** (260 mg, 0.95 mmol) was dissolved in MeOH (7.5 mL). A solution of LiOH·H₂O (200 mg, 4.76 mmol) in H₂O (2.5 mL) was then added and the mixture stirred for 6 h. The reaction was acidified with HCl (1 N) until pH 2 and then extracted with EtOAc (3 x 10 mL). The organic fractions were combined and washed with H₂O and brine, dried over MgSO₄, and filtered. Evaporation under reduced pressure left the product pure **46** as a brown solid (200 mg, 0.77 mmol, 81%). TLC R_f 0.13 (40% EtOAc/hexanes); ¹H-NMR (300 MHz, CD₃OD) δ 7.47 (t, 2H), 7.41 (d, 1H), 7.28 (d, 2H), 7.19 (dd, 1H), 6.18 (dd, 1H), 6.05 (m, 1H), 3.27 (s, 3H), 3.40 (m, 2H), 2.22 (t, 2H).

(E,E)-Octa-2,4-dienedioic acid 1-hydroxyamide 8-phenylamide (47)



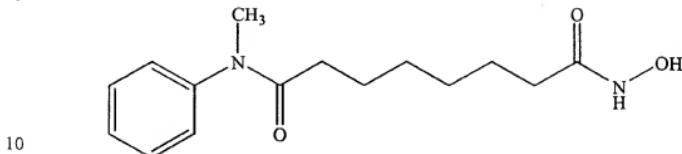
Acid **46** (200 mg, 0.77 mmol) and TBDPSSO-NH₂ (220 mg, 0.81 mmol) were dissolved in CH₂Cl₂ (8 mL). To this stirred solution were added EDC (178 mg, 0.93 mmol) and DMAP (5 mg) and the reaction allowed to proceed overnight. The mixture was concentrated and 25 then passed through a plug of silica gel (EtOAc). Evaporation under reduced pressure left a light brown oil (383 mg, 0.75 mmol, 97%). The protected hydroxamate (270 mg, 0.53 mmol) was dissolved in CH₂Cl₂ (10 mL) and TFA was added (0.5 mL). The solution was stirred for 2 h, and a new spot on TLC was observed 30 which stained with FeCl₃. The solution was concentrated under reduced pressure and diethyl ether added, giving a residue which adhered to the flask. The liquid phase was drawn off, the residue was triturated with EtOAc, the liquid removed, and evaporation of all volatiles from the residue gave **47** as a brown 35 gum (23 mg, 0.084 mmol, 16%). TLC R_f 0.22 (5% MeOH/CH₂Cl₂); ¹H-NMR (400 MHz, CD₃OD) δ 7.50 (t, 2H), 7.40 (t, 1H), 2.27 (d, 2H),

-69-

7.08 (m, 1H), 6.11 (m, 1H), 5.97 (m, 1H), 5.80 (m, 1H), 3.23 (s, 3H), 3.39 (m, 2H), 2.21 (t, 2H).

Octanedioic acid hydroxyamide phenylamide (48)

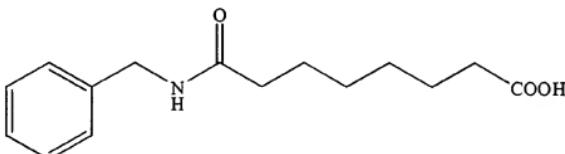
5



The title compound **48** was obtained as a brown gum (9 mg) by a series of steps analogous to the preparation of **47**. TLC R_f 0.20 (5% MeOH/CH₂Cl₂); ¹H-NMR (400 MHz, CD₃OD) δ 7.51 (t, 2H), 7.41 (t, 1H), 7.30 (d, 2H), 3.29 (s, 3H), 2.11 (m, 4H), 1.58 (m, 4H), 1.22 (m, 4H).

Octanedioic acid benzylamide (49)

20



25

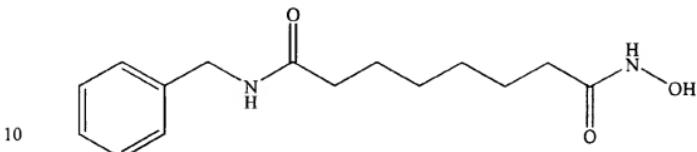
To a stirred solution of suberoyl chloride (1.00 mL, 5.55 mmol) in THF (40 mL) at 0 °C was added a solution of benzylamine (0.61 mL, 5.55 mmol) and DIEA (1.45 mL, 8.33 mmol) in THF (10 mL) dropwise. The mixture was allowed to warm to ambient temperature and stirred for 1 h. Then, HCl (10 mL, 1 N) was added and the mixture stirred for 0.5 h. The contents were diluted with EtOAc (30 mL) and the layers separated. The aqueous portion was extracted with EtOAc (3 x 10mL), the organics combined, washed with brine (5 mL), and dried over MgSO₄. Filtration and concentration under reduced pressure left **49** as an off-white solid. ¹H-NMR (300 MHz, DMSO-*d*₆) δ 11.98 (br

-70-

s, 1H), 9.80 (t, 1H), 7.32 (m, 2H), 7.23 (m, 3H), 4.25 (d, 2H), 2.19 (t, 2H), 2.12 (t, 2H), 1.50 (m, 4H), 1.25 (m, 4H).

Octanedioic acid benzylamide hydroxyamide (50)

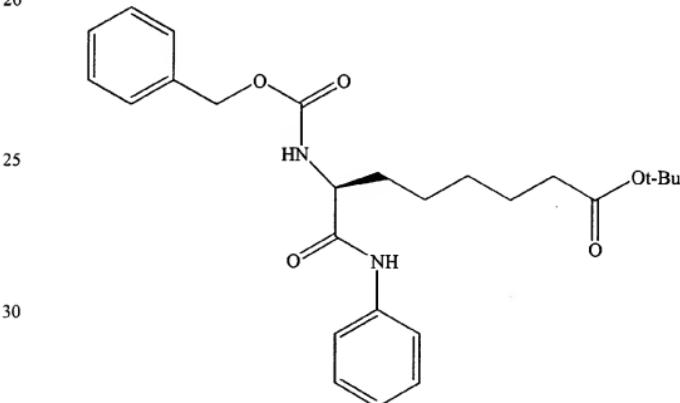
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This compound was prepared from **49** through its protected hydroxamate as described for earlier compounds. Obtained **50** as a white solid. ¹H-NMR (400 MHz, DMSO-*d*₆) δ 10.30 (s, 1H), 8.27 (t, 1H), 7.28 (m, 2H), 7.23 (m, 3H), 5.65 (d, 2H), 2.11 (t, 2H), 1.91 (t, 2H), 1.46 (m, 4H), 1.23 (m, 4H).

(7*S*)-7-Benzoyloxycarbonylamino-7-phenylcarbamoyl-heptanoic acid *t*-butyl ester (51)

20

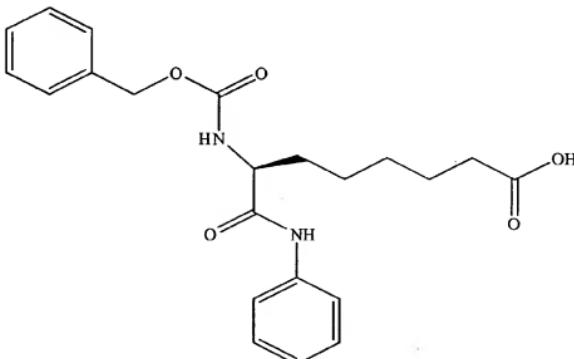


35 *N*-Cbz-L-2-aminosuberic acid 8-*t*-butyl ester, dicyclohexylamine salt (100 mg, 0.18 mmol) was dissolved in HCl (5 mL, 1 N) and

extracted with EtOAc (3 x 10 mL). The extracts were combined, washed with brine, and dried over MgSO₄. Evaporation left the free acid as a white solid (68 mg, 0.179 mmol). This was dissolved in CH₂Cl₂ (2.5 mL), to which were added aniline (17 μ L, 0.19 mmol), DIEA (46 μ L, 0.27 mmol), and finally Py-BOP (97 mg, 0.19 mmol). The solution was stirred for 1 h, then concentrated, and the residue partitioned between H₂O (5 mL) and EtOAc (10 mL). The layers were separated, and the aqueous portion extracted with EtOAc (3 x 10 mL). The extracts were pooled and washed with HCl (1 N), then brine, dried over MgSO₄, and filtered. Concentration under reduced pressure gave a solid residue which was passed through a plug of silica gel (30% EtOAc/hexanes). The collected eluent was evaporated to give 51 as a white solid (76 mg, 0.167 mmol, 94%). TLC R_f 0.38 (30% EtOAc/hexanes); ¹H-NMR (400 MHz, CDCl₃) δ 8.21 (s, 1H), 7.48 (d, 2H), 7.32 (m, 5 H), 7.28 (t, 2H), 7.08 (t, 1H), 5.39 (br d, 1H), 5.10 (m, 2H), 4.26 (br dd, 1H), 2.07 (t, 2H), 1.92 (m, 1H), 1.66 (m, 1H), 1.55 (m, 2H), 1.42 (s, 9H), 1.38 (m, 4H).

20 (7S)-7-Benzylloxycarbonylamino-7-phenylcarbamoyl-heptanoic acid (52)

25



30

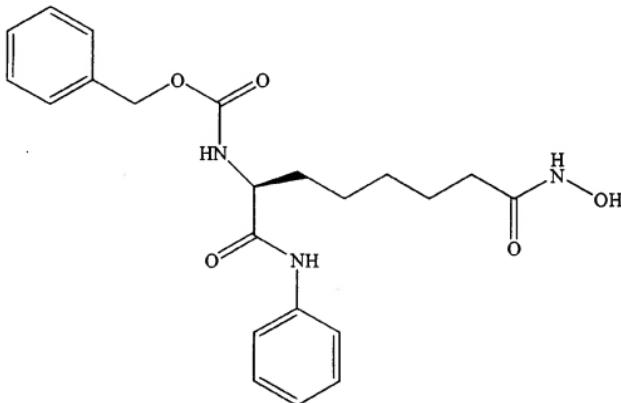
35 To a solution of ester 51 (76 mg, 0.167 mmol) in CH₂Cl₂ (5 mL) was added TFA (0.5 mL) and the reaction solution stirred for 5

h. The solution was concentrated under reduced pressure to give crude **52** as a white solid (80 mg) which was used in the next step without purification. TLC R_f 0.32 (5% MeOH/CH₂Cl₂); ¹H-NMR (400 MHz, DMSO-*d*₆) δ 11.93 (br s, 1H), 9.99 (s, 1H), 7.58 (d, 5 2H), 7.55 (d, 1H), 7.35 (m, 4H), 7.29 (t, 2H), 7.03 (t, 1H), 5.02 (m, 2H), 4.11 (br dd, 1H), 2.17 (t, 2H), 1.59 (m, 2H), 1.48 (m, 2H), 1.22 (m, 4H).

(1*S*)-(6-Hydroxycarbamoyl-1-phenylcarbamoyl-hexyl)-carbamic acid 10 benzyl ester (53)

15

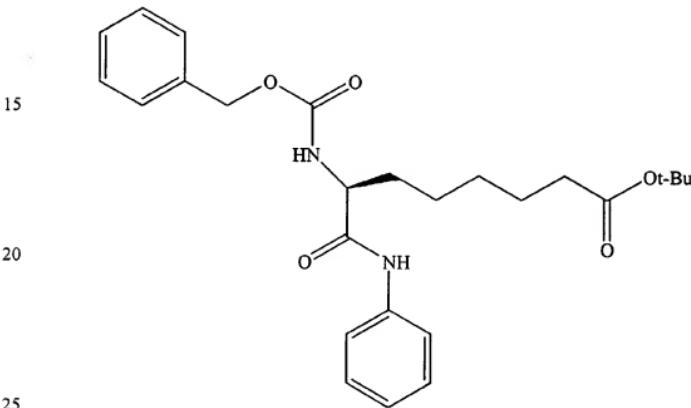
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25 To a solution of crude acid **52** (80 mg) and TB DPSO-NH₂ (60 mg, 0.221 mmol) in CH₂Cl₂ were added DIEA (52 μ L, 0.302 mmol) followed by Py-BOP (125 mg, 0.241 mmol). The solution was stirred for 3 h, then concentrated under reduced pressure. The residue was passed through a plug of silica gel (50% 30 EtOAc/hexanes) and the collected eluent evaporated. A white foam (107 mg, 0.164 mmol, 82%) was obtained, this was dissolved in CH₂Cl₂ (5 mL) and TFA (0.25 mL) was added and the solution stirred for 2 h. A new spot that stained with FeCl₃ was indicated by TLC analysis. The mixture was concentrated under 35 reduced pressure, and the residue was solvated in a minimum of EtOAc and the product precipitated with hexanes. The resulting

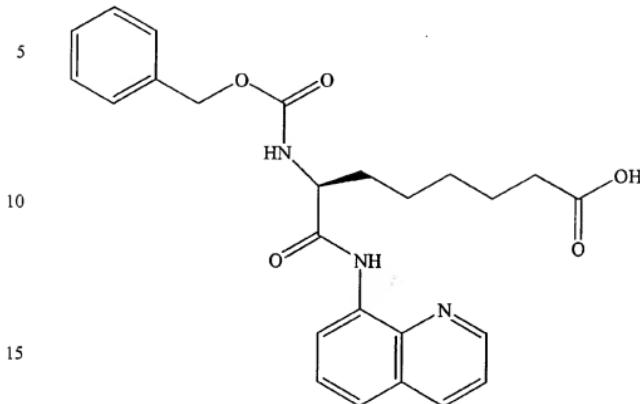
white gel was rinsed with hexanes and dried under vacuum, to give **53** as a white solid (40 mg, 0.097 mmol, 58% over three steps). ¹H-NMR (400 MHz, DMSO-*d*₆) δ 10.31 (s, 1H), 9.99 (s, 1H), 7.59 (d, 2H), 7.56 (d, 1H), 7.37 (m, 4H), 7.29 (t, 2H), 7.02 5 (t, 1H), 5.02 (m, 2H), 4.11 (dt, 1H), 1.90 (t, 2H), 1.61 (m, 2H), 1.47 (m, 2H), 1.30 (m, 4H). MS (ESI+) calcd for C₂₂H₂₇N₃O₅ 413, found 414 [M+H]⁺.

(7*S*)-7-Benzylloxycarbonylamino-7-(quinolin-8-ylcarbamoyl)-10 heptanoic acid *t*-butyl ester (**54**)



The title compound was made from *N*-Cbz-L-2-aminosuberic acid 8-*t*-butyl ester, dicyclohexylamine salt in a manner similar to that for **51**. Flash chromatography (0-1% MeOH/CH₂Cl₂) gave **54** as a light brown solid (70 mg, 0.138 mmol, 82%). TLC R_f 0.42 (2% 30 MeOH/CH₂Cl₂); ¹H-NMR (400 MHz, CDCl₃) δ 10.19 (s, 1H), 8.77 (dd, 1H), 8.71 (dd, 1H), 8.15 (dd, 1H), 7.52 (m, 2H), 7.45 (m, 1H), 7.33 (m, 4H), 5.50 (br d, 1H), 5.15 (m, 2H), 4.51 (br dd, 1H), 2.17 (t, 2H), 2.00 (m, 1H), 1.79 (m, 1H), 1.56 (m, 2H), 1.45 (m, 2H), 1.40 (s, 9H), 1.38 (m, 2H).

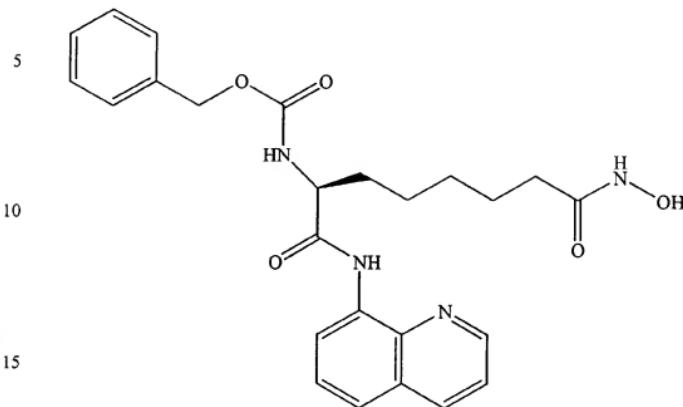
(7*S*)-7-Benzylloxycarbonylamino-7-(quinolin-8-ylcarbamoyl)-
heptanoic acid (55)



Prepared from **54** in a manner similar to that for **52**. Obtained **55** as a brown solid (72 mg, 0.129 mmol). TLC R_f 0.16 (50% EtOAc/hexanes); $^1\text{H-NMR}$ (400 MHz, $\text{DMSO-}d_6$) δ 11.92 (br s, 1H), 10.46 (s, 1H), 8.49 (dd, 1H), 8.63 (dd, 1H), 8.42 (dd, 1H), 8.10 (d, 1H), 7.68 (dd, 1H), 7.58 (t, 1H), 7.36 (m, 2H), 7.28 (m, 2H), 5.09 (m, 2H), 4.22 (m, 1H), 2.19 (t, 2H), 1.83 (m, 1H), 1.67 (m, 1H), 1.48 (m, 2H), 1.39 (m, 2H), 1.28 (m, 2H).

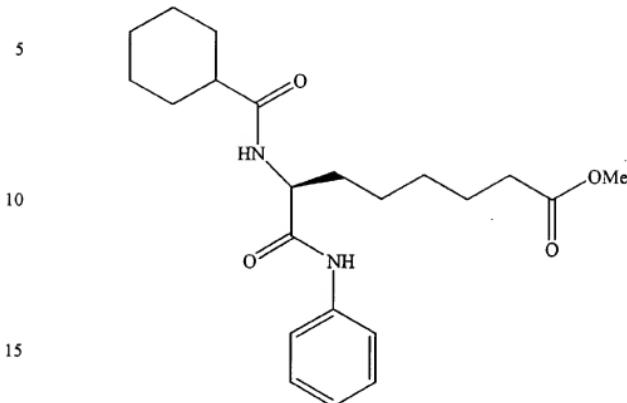
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(1*S*) - [6-Hydroxycarbamoyl-1-(quinolin-8-ylcarbamoyl)-hexyl]-carbamic acid benzyl ester (56)



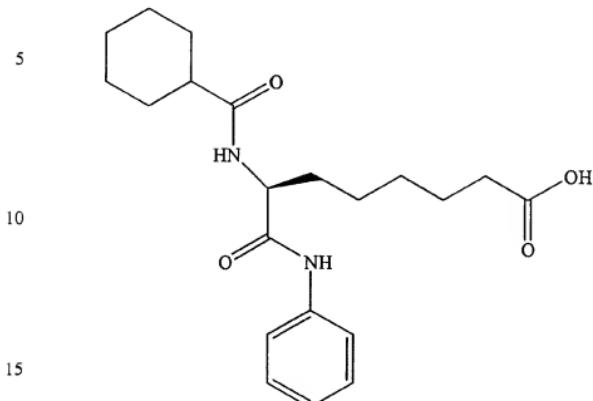
Prepared from 55 in a manner similar to that for 53. Obtained 56 as a white solid (15 mg, 0.032 mmol, 44%). ¹H-NMR (400 MHz, DMSO-*d*₆) δ 10.46 (s, 1H), 10.31 (s, 1H), 8.85 (dd, 1H), 8.63 (dd, 1H), 8.42 (dd, 1H), 8.12 (d, 1H), 8.66 (m, 2H), 7.58 (t, 1H), 7.37 (m, 2H), 7.28 (m, 2H), 7.20-6.90 (1H), 5.10 (m, 2H), 4.10 (m, 1H), 1.92 (t, 2H), 1.82 (m, 1H), 1.68 (m, 1H), 1.49 (m, 2H), 1.40 (m, 2H), 1.26 (m, 2H). MS (ESI+) calcd for C₂₅H₂₈N₄O₅ 464, found 465 [M+H]⁺.

(7S)- (Cyclohexanecarbonyl-amino)-7-phenylcarbamoyl-heptanoic acid methyl ester (57)



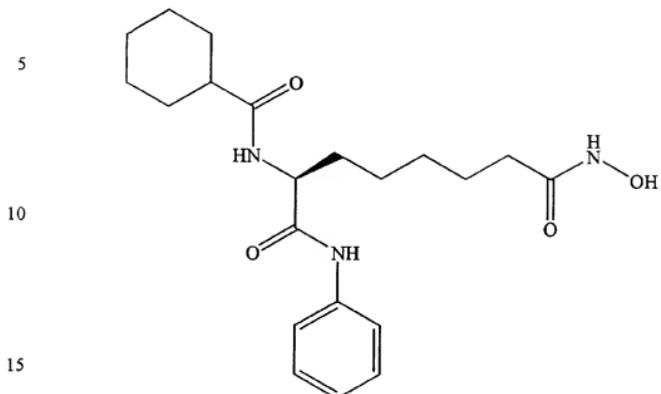
To a solution of **5** (81 mg, 0.214 mmol) in CH_2Cl_2 (10 mL) was added TFA (0.5 mL) and the solution stirred for 2 h. The mixture was concentrated under reduced pressure. To a solution of this amine (62 mg, 0.223 mmol) and cyclohexane carboxylic acid (31 μL , 0.245 mmol) in CH_2Cl_2 (4 mL) were added Py•BOP (140 mg, 0.268 mmol) and DIIEA (58 μL , 0.335 mmol). The solution was stirred for 2 h, concentrated under reduced pressure, and the product purified by flash chromatography (40% EtOAc/hexanes). Evaporation left crude **57** as a white solid (95 mg) containing a small amount of unreacted cyclohexane acid impurity. This material was used in the next step without further purification. TLC R_f 0.58 (50% EtOAc/hexanes); $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 8.58 (s, 1H), 7.50 (d, 2H), 7.28 (t, 2H), 7.07 (t, 1H), 6.14 (d, 1H), 4.56 (dt, 1H), 3.64 (s, 3H), 2.28 (t, 2H), 2.13 (tt, 1H), 1.94 (m, 1H), 1.85 (m, 2H), 1.76 (m, 2H), 1.64 (m, 4H), 1.41 (m, 5H), 1.22 (m, 4H).

(7*S*)-(Cyclohexanecarbonyl-amino)-7-phenylcarbamoyl-heptanoic acid (58)



To a solution of ester **57** (95 mg) in MeOH (2.5 mL) at 0 °C was added a solution of NaOH (1 M, 2.5 mL). A white precipitate 20 formed upon addition, which was re-dissolved by the addition of THF (2.5 mL). Additional NaOH (1 M, 1.0 mL) was added after 3 h and the temperature maintained at 0 °C. Upon complete disappearance of starting material by TLC analysis, the reaction 25 contents were acidified with HCl (1 N) to obtain a white precipitate. The supernatant was drawn off, and the solid filtered under aspiration. The combined liquors were extracted with EtOAc (3 x 5 mL), and the extracts combined, washed with brine, dried over MgSO₄, and filtered. Concentration under 30 reduced pressure left a white solid which was combined with the filter cake and dried under vacuum to obtain the carboxylic acid **58** (75 mg, 0.200 mmol, 90%). ¹H-NMR (400 MHz, DMSO-*d*₆) δ 11.95 (s, 1H), 9.98 (s, 1H), 7.90 (d, 1H), 7.58 (d, 1H), 7.28 (t, 2H), 7.02 (t, 1H), 4.33 (dt, 1H), 2.22 (tt, 1H), 2.17 (t, 2H), 1.67 (m, 6H), 1.60 (m, 2H), 1.46 (m, 2H), 1.22 (m, 9H).

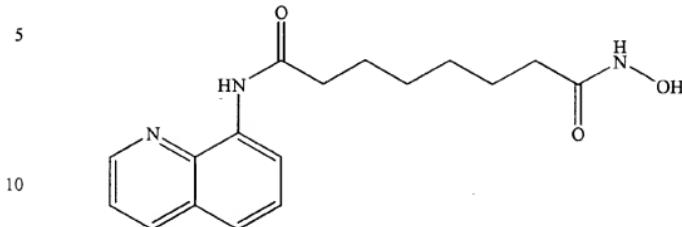
(2S)-2-(Cyclohexanecarbonyl-amino)-octanedioic acid 8-
hydroxyamide 1-phenylamide (59)



Acid 58 (70 mg, 0.187 mmol), TBDDPSO-NH₂ (61 mg, 0.224 mmol), and DMAP (5 mg) were dissolved in CH₂Cl₂ (4 mL) and EDC (47 mg, 0.243 mmol) was added. The solution was stirred overnight. After 20 concentration under reduced pressure, the material was purified by flash chromatography (50% EtOAc/hexanes). Evaporation of the combined product fractions gave a white foam (80 mg, 0.131 mmol, 70%). To a solution of this protected hydroxamate in CH₂Cl₂ (2 mL) and THF (3 mL) was added TFA (0.25 mL) and stirred for 1.5 h. A new spot which stained immediately with FeCl₃ was observed on TLC. The solution was concentrated and all volatiles removed under vacuum. The residue was triturated with EtOAc and obtain a white gel precipitate which was transferred to a plastic tube with EtOAc (5 mL). The tube was centrifuged to form a pellet, 30 the supernatant drained, and EtOAc (10 mL) added. The pellet was resuspended with sonication, then centrifuged again, the supernatant discarded, and the residue dried under vacuum. A white solid 59 (18 mg, 0.046 mmol, 35%) was obtained. ¹H-NMR (400 MHz, DMSO-*d*₆) δ 10.31 (s, 1H), 9.97 (s, 1H), 7.89 (d, 1H), 7.57 (d, 2H), 7.28 (t, 2H), 7.02 (t, 1H), 4.33 (dt, 1H), 2.22 (t, 2H), 1.91 (t, 2H), 1.61 (m, 6H), 1.68 (m, 2H), 1.45 (m, 2H),

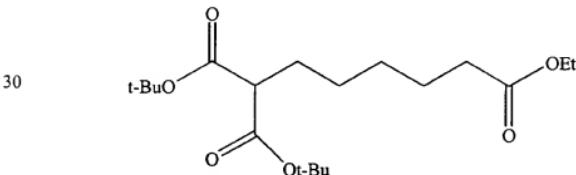
1.21 (9H).

Octanedioic acid hydroxyamide quinolin-8-ylamide (60)



This compound was prepared from suberic acid monomethyl ester in similar fashion to **48**, with the use of 8-aminoquinoline. The 15 crude residue obtained after TFA deprotection of the protected hydroxamate was taken up in a small volume of EtOAc and precipitated with hexanes to give **60** as a white solid (18 mg, 0.057 mmol, 21% from the carboxylic acid). ¹H-NMR (400 MHz, DMSO-*d*₆) δ 10.31 (s, 1H), 10.02 (s, 1H), 8.92 (dd, 1H), 8.61 (dd, 1H), 8.40 (dd, 1H), 7.65 (dd, 1H), 7.63 (dd, 1H), 7.56 (t, 1H), 2.56 (t, 1H), 1.93 (t, 1H), 1.63 (m, 2H), 1.49 (m, 2H), 1.28 (m, 4H). MS (ESI+) calcd for C₁₇H₂₁N₃O₃ 315, found 316 [M+H]⁺.

25 **2-*t*-Butoxycarbonyl-octanedioic acid 1-*t*-butyl ester 8-ethyl ester (61)**

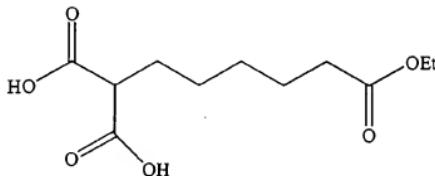


To a stirred suspension of NaH (60% disp., 197 mg, 4.913 mmol) 35 in THF (25 mL) at 0 °C was added di-*t*-butyl malonate (1.00 mL, 4.466 mmol) and the mixture allowed to warm to ambient

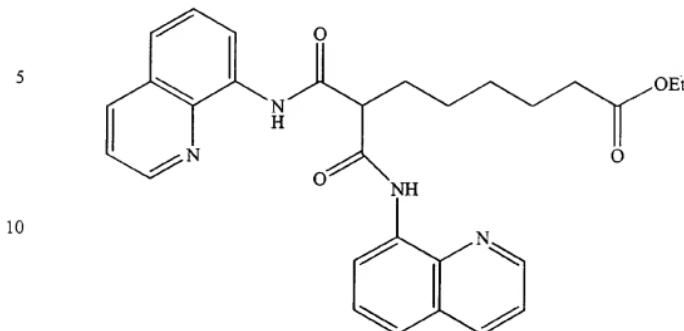
temperature. After 1 h, gas had ceased evolving and ethyl 6-bromohexanoate (0.88 mL, 4.913 mmol) was added dropwise. The reaction was brought to reflux overnight. The reaction was carefully quenched with H₂O (10 mL) and diluted with EtOAc. 5 After separation of the layers, the aqueous portion was extracted with EtOAc (3 x 10 mL). The extracts were pooled and washed with H₂O, then brine, dried over MgSO₄, and filtered. Concentration under reduced pressure gave a yellow oil which was passed through a plug of silica gel (10% EtOAc/hexanes). 10 Evaporation left a light yellow syrup **61** (1.52 g, 4.24 mmol, 95%). TLC R_f 0.44 (10% EtOAc/hexanes); ¹H-NMR (400 MHz, CDCl₃) δ 4.10 (q, 2H), 3.08 (t, 1H), 2.26 (t, 2H), 1.76 (m, 2H), 1.60 (m, 2H), 1.43 (s, 18H), 1.32 (m, 4H), 1.23 (m, 3H).

15 **2-Carboxy-octanedioic acid 8-ethyl ester (62)**

20

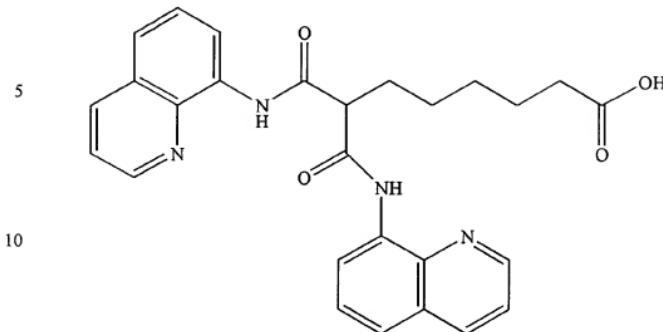


To a solution of triester **61** (500 mg, 1.395 mmol) in CH₂Cl₂ (20 25 mL) was added TFA (2.0 mL) and the reaction mixture stirred overnight. Volatile components were evaporated under vacuum, and the residue repeatedly dissolved in CH₂Cl₂ and evaporated to remove all traces of TFA. A solid **62** (327 mg, 1.33 mmol) was obtained and used directly in the next step without further 30 purification. ¹H-NMR (400 MHz, DMSO-*d*₆) δ 12.62 (br s, 2H), 4.03 (q, 2H), 3.16 (t, 1H), 2.25 (t, 2H), 1.67 (m, 2H), 1.49 (m, 2H), 1.25 (m, 4H), 1.16 (t, 3H).

7,7-Bis-(quinolin-8-ylcarbamoyl)-heptanoic acid ethyl ester (65)

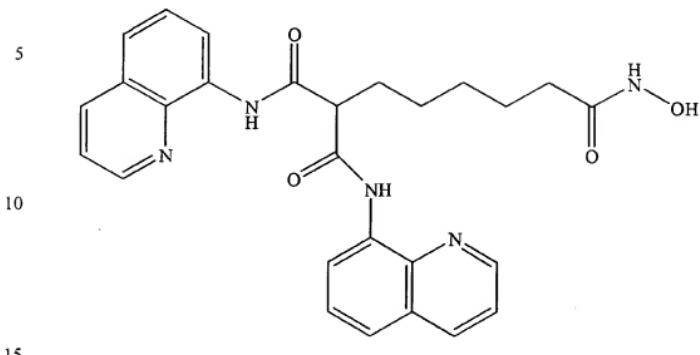
15 Diacid **62** (150 mg, 0.609 mmol), 8-aminoquinoline (211 mg, 1.462 mmol), and DMAP (5 mg) were dissolved in THF (6 mL). To this solution was added EDC (350 mg, 1.827 mmol) and the reaction allowed to proceed overnight. The mixture was concentrated under reduced pressure and the product purified by flash 20 chromatography (40% EtOAc/hexanes). Evaporation of the combined product fractions left **63** as a light brown solid (100 mg, 0.201 mmol, 14%). ¹H-NMR (400 MHz, DMSO-*d*₆) δ 10.85 (s, 2H), 8.92 (dd, 2H), 8.64 (dd, 2H), 8.40 (dd, 2H), 7.68 (dd, 2H), 7.62 (dd, 2H), 7.57 (t, 2H), 4.35 (t, 1H), 3.98 (q, 2H), 2.24 (t, 2H), 2.00 (m, 2H), 1.51 (m, 2H), 1.37 (m, 4H), 1.12 (t, 3H).

7,7-Bis-(quinolin-8-ylcarbamoyl)-heptanoic acid (64)



15 To a solution of ester **63** (94 mg, 0.212 mmol) in MeOH (3 mL) and THF (1 mL) was added a solution of LiOH· H₂O (44 mg, 1.062 mmol) in H₂O (1 mL) and the mixture was stirred for 5 h. After acidification with HCl (1 N) to pH 7, EtOAc (10 mL) was added and the layers separated. The aqueous portion was extracted 20 with EtOAc (3 x 5 mL), and the extracts combined, washed with sat. NH₄Cl (3 mL), H₂O (3 mL), then brine, dried over MgSO₄, and filtered. Concentration under reduced pressure left **64** as a white solid (94 mg, 0.200 mmol, 94%). TLC R_f 0.21 (50% EtOAc/hexanes); ¹H-NMR (400 MHz, DMSO-d₆) δ 11.88 (s, 1H), 10.85 25 (s, 2H), 8.93 (dd, 2H), 8.65 (dd, 2H), 8.40 (dd, 2H), 7.69 (dd, 2H), 7.63 (dd, 2H), 7.58 (t, 2H), 4.35 (t, 1H), 2.16 (t, 2H), 2.00 (m, 2H), 1.49 (m, 2H), 1.38 (m, 4H).

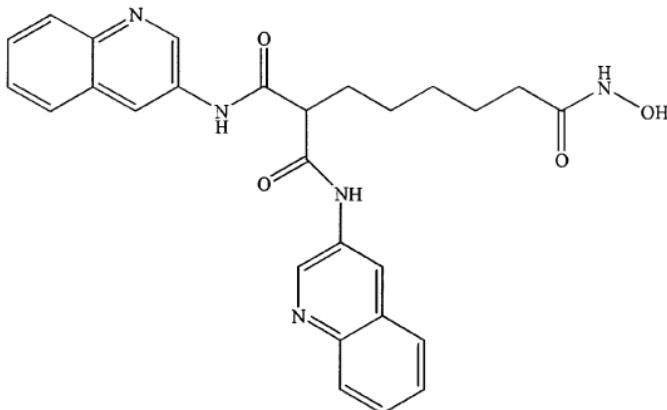
2-(Quinolin-8-ylcarbamoyl)-octanedioic acid 8-hydroxyamide 1-
quinolin-8-ylamide (65)



15

Acid **64** (94 mg, 0.200 mmol), TBDPSSO-NH₂ (74 mg, 0.272 mmol), and DMAP (5 mg) were dissolved in CH₂Cl₂ (4 mL) and EDC (57 mg, 0.295 mmol) was added. The solution was stirred overnight, then concentrated under reduced pressure. Purification by flash chromatography (30-50% EtOAc/hexanes) and evaporation of the combined product fractions gave a white foam. To a solution of this protected hydroxamate in CH₂Cl₂ (4 mL) was added TFA (0.2 mL) and the solution stirred for 4 h. TLC indicated complete consumption of starting material and a new spot that stained 20 with FeCl₃. The solution was concentrated under reduced pressure, and the residue dissolved in a minimum of EtOAc. Addition of hexanes gave a white precipitate, from which the mother liquor was removed. After rinsing with hexanes, the residue was dried under vacuum to leave **65** as a white solid (30 25 mg, 0.061 mmol, 22% from the carboxylic acid). ¹H-NMR (400 MHz, CDCl₃) δ 10.85 (s, 2H), 10.30 (s, 1H), 8.93 (dd, 2H), 8.65 (dd, 2H), 8.40 (dd, 2H), 7.69 (dd, 2H), 7.63 (dd, 2H), 7.58 (t, 2H), 4.35 (t, 1H), 1.99 (m, 2H), 1.92 (t, 2H), 1.48 (m, 2H), 1.35 (m, 4H). MS (ESI+) calcd for C₂₇H₂₇N₅O₄ 485, found 486 [M+H]⁺.

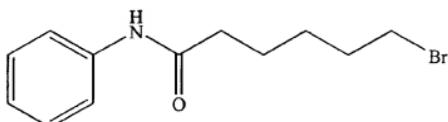
**2-(Quinolin-3-ylcarbamoyl)-octanedioic acid 8-hydroxyamide 1-
quinolin-3-ylamide (68)**



The title compound was made from diacid **62** as analogous to **65**.
 5 ¹H-NMR (400 MHz, DMSO-*d*₆) δ 10.60 (s, 1H), 10.34 (s, 1H), 8.95 (dd, 2H), 8.74 (s, 2H), 7.93 (dd, 2H), 7.64 (dd, 2H), 7.56 (dd, 2H), 3.71 (t, 1H), 1.96 (m, 4H), 1.51 (m, 2H), 1.34 (m, 4H).

6-Bromohexanoic acid phenylamide (76)

10



15

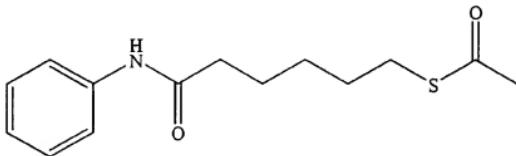
To a solution of 6-bromohexanoyl chloride (1.00 mL, 6.53 mmol) in THF (35 mL) at 0 °C was added dropwise a solution of aniline (0.60 mL, 6.53 mmol) and TEA (1.09 mL, 7.84 mmol) in THF (5 mL). The reaction mixture was allowed to warm to ambient temperature 20 and stirred for 2 h. The mixture was filtered, the solids rinsed with EtOAc, and the filtrate reduced under vacuum. The

residue was partitioned between H_2O (15 mL) and EtOAc (20 mL) and the layers separated. The aqueous portion was extracted with EtOAc (3 x 10 mL) and the organic layers combined, washed with HCl (1 N), brine, dried over $MgSO_4$, and filtered. 5 Concentration under reduced pressure left a brown oil which was passed through a plug of silica gel (30% EtOAc/hexanes) under aspiration. Concentration under reduced pressure left 67 as a solid (1.55 g, 5.74 mmol, 88%). TLC R_f 0.36 (25% EtOAc/hexanes); 1H -NMR (400 MHz, $DMSO-d_6$) δ 9.85 (s, 1H), 7.57 10 (d, 2H), 7.27 (t, 2H), 7.01 (t, 1H), 3.53 (t, 2H), 2.30 (t, 2H), 1.81 (t, 2H), 1.63 (m, 2H), 1.42 (m, 2H); MS (ESI+) calcd for $C_{12}H_{16}BrNO$ 268+270, found 269+271 [M+H]⁺.

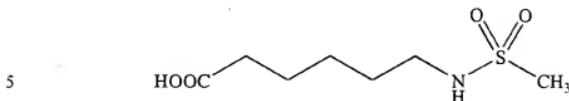
Thioacetic acid S-(5-phenylcarbamoyl-pentyl) ester (68)

15

20

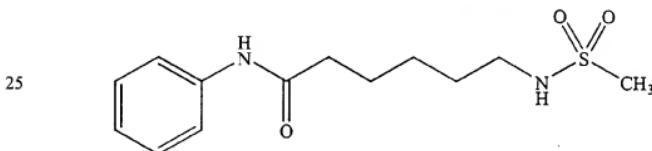


Bromide 67 (200 mg, 0.74 mmol), potassium thioacetate (110 mg, 0.96 mmol), and sodium iodide (10 mg) were combined in THF (6 25 mL) and the vigorously stirred mixture brought to reflux overnight. The reaction mixture was concentrated, the passed through a plug of silica gel (20% EtOAc/hexanes, 200 mL) under aspiration. Evaporation under reduced pressure left 68 as an orange crystalline solid (190 mg, 0.72 mmol, 97%). TLC R_f 0.22 30 (25% EtOAc/hexanes); 1H -NMR (400 MHz, $DMSO-d_6$) δ 9.83 (s, 1H), 7.56 (d, 2H), 7.27 (t, 2H), 7.00 (t, 1H), 2.82 (t, 2H), 2.30 (s, 3H), 2.28 (t, 2H), 1.57 (m, 2H), 1.52 (m, 2H), 1.35 (m, 2H).

6-Methanesulfonylamino-hexanoic acid (69)

6-aminohexanoic acid (904 mg, 6.89 mmol) and NaOH (415 mg, 10.34 mmol) were dissolved in H₂O (30 mL) and cooled to 0-5 °C. 10 Methanesulfonyl chloride (0.586 mL, 7.58 mmol) was added dropwise and the reaction mixture stirred for 2 h, then warmed to ambient temperature and stirred for an additional 2 h. The mixture was acidified with HCl (1 N) and extracted with EtOAc (3 x 15 mL). The extracts were combined, washed with H₂O, then 15 brine, dried over MgSO₄, and filtered. Evaporation under reduced pressure gave **69** as a white crystalline solid (207 mg, 0.99 mmol, 14%). ¹H-NMR (400 MHz, DMSO-d₆) δ 11.95 (s, 1H), 6.91 (t, 1H), 2.90 (dt, 2H), 2.87 (s, 3H), 2.20 (t, 2H), 2.48 (m, 2H), 2.43 (m, 2H), 1.27 (m, 2H).

20

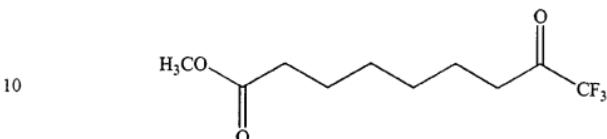
6-Methanesulfonylamino-hexanoic acid phenylamide (70)

To a solution of acid **69** (100 mg, 0.48 mmol), aniline (60 μL, 30 0.66 mmol), and DMAP (5 mg) in THF (5 mL) was added EDC (119 mg, 0.57 mmol). The reaction mixture was stirred overnight, then partitioned between H₂O (10 mL) and EtOAc (15 mL). The layers were separated, and the aqueous portion extracted with EtOAc (3 x 10 mL). The organic fractions were combined, washed with sat. 35 NH₄Cl (5 mL), then brine, dried over MgSO₄, and filtered. Concentration under reduced pressure gave **70** as a white

crystalline solid (130 mg, 0.46 mmol, 95%). ¹H-NMR (400 MHz, DMSO-*d*₆) δ 9.84 (s, 1H), 7.57 (d, 2H), 7.26 (t, 2H), 7.00 (t, 1H), 6.92 (t, 1H), 2.91 (dt, 2H), 2.85 (s, 3H), 1.58 (m, 2H), 1.47 (m, 2H), 1.31 (m, 2H).

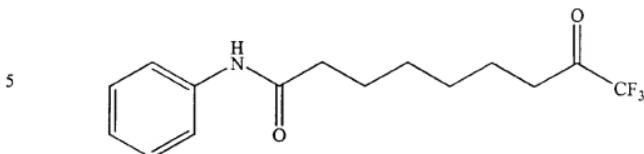
5

9,9,9-trifluoro-8-oxononanoic acid methyl ester (71)



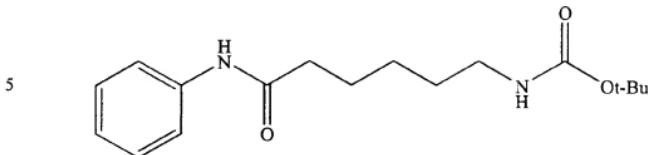
To a solution of suberic acid monomethyl ester (1.00 g, 5.31 mmol) in THF (15 mL) was added oxalyl chloride (2 mL) followed 15 by DMF (1 drop). The solution was stirred for 2 h, then concentrated under reduced pressure. Volatiles were removed under high vacuum overnight, leaving a yellow oil (1.08 g, 5.22 mmol, 98%). This crude acid chloride was then transformed into the trifluoromethyl ketone by a literature method as follows. 20 (65) To a solution of the acid chloride (1.08 g, 5.22 mmol) in CH₂Cl₂ (45 mL) at 0 °C were added trifluoroacetic anhydride (4.64 mL, 32.81 mmol) and pyridine (3.54 mL, 43.74 mmol). The mixture was allowed to warm to ambient temperature and stirred for 2 h. After returning to 0 °C, ice-cold H₂O (20 mL) was added 25 carefully. Additional H₂O (100 mL) was added and the layers separated. The aqueous phase was extracted with CH₂Cl₂ (2 x 30 mL) and the organic layers combined, washed with brine, dried over MgSO₄, and filtered. Evaporation under reduced pressure left a brown oil, which was purified by flash chromatography (2-30 4% MeOH/CH₂Cl₂) to give 71 as a clear oil (641 mg, 2.67 mmol, 49%). TLC R_f 0.24 (2% MeOH/CH₂Cl₂); ¹H-NMR (400 MHz, CDCl₃) δ 3.67 (s, 3H), 2.71 (t, 2H), 2.31 (t, 2H), 1.65 (m, 4H), 1.35 (m, 4H).

9,9,9-Trifluoro-8-oxo-nonanoic acid phenylamide (72)



To a solution of ester 71 (300 mg, 1.25 mmol) in THF (18 mL) was 10 added a solution of LiOH• H₂O (262 mg, 6.24 mmol) in H₂O (6 mL) and the suspension was stirred overnight. The mixture was then acidified with HCl (1 N) to pH 2 and then extracted with EtOAc (3 x 15 mL). The extracts were combined, washed with brine, dried over MgSO₄, and filtered. Concentration under reduced 15 pressure left a white solid (211 mg, 0.93 mmol, 75%). To a solution of this acid (109 mg, 0.48 mmol), EDC (111 mg, 0.58 mmol), and DMAP (5 mg) in CH₂Cl₂ (5 mL) was added aniline (49 μ L, 0.53 mmol) and the reaction allowed to proceed overnight. The solution was partitioned between H₂O (5 mL) and EtOAc (10 mL). 20 The layers were separated, and the aqueous phase extracted with EtOAc (3 x 5 mL). The organic portions were combined, washed with brine, dried over MgSO₄, and filtered. Evaporation under reduced pressure left a solid which was purified by preparative TLC (30% EtOAc/hexanes) with isolation of the least polar band 25 by EtOAc extraction. The extract was concentrated to give 72 as a yellowish solid (92 mg, 0.31 mmol, 65%). TLC R_f 0.48 (50% EtOAc/hexanes); ¹H-NMR (400 MHz, CDCl₃) δ 7.51 (d, 2H), 7.32 (t, 2H), 7.10 (t, 1H), 2.72 (t, 2H), 2.36 (t, 2H), 1.72 (m, 4H), 1.40 (m, 4H); ¹⁹F NMR (? MHz, CDCl₃) -78.40 (s, 3F); MS (APCI+) 30 calcd for C₁₅H₁₉F₃NO₂ 301, found 325 [M+Na]⁺.

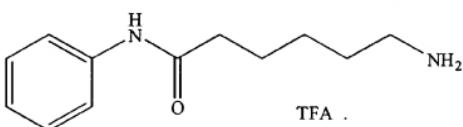
(5-Phenylcarbamoyl-pentylyl)-carbamic acid t-butyl ester (73)



To a solution of *N*-Boc-6-aminohexanoic acid (2.50 g, 10.81 10 mmol), EDC (2.69 g, 14.05 mmol), and DMAP (20 mg) in CH_2Cl_2 (100 mL) was added aniline (1.04 mL, 11.35 mmol) and the mixture stirred overnight. The solution was evaporated under reduced pressure to a small volume, then partitioned between H_2O (20 mL) and EtOAc (30 mL). The layers were separated, and the aqueous 15 phase extracted with EtOAc (3 x 15 mL). The organic portions were combined, washed with sat. NH_4Cl (5 mL), then brine, dried over MgSO_4 , and filtered. Concentration under reduced pressure left pure 73 as a white solid (3.14 g, 10.25 mmol, 95%). TLC R_f 0.40 (50% EtOAc/hexanes); $^1\text{H-NMR}$ (400 MHz, $\text{DMSO}-d_6$) δ 9.81 (s, 20 1H), 7.56 (d, 2H), 7.26 (t, 2H), 7.00 (t, 1H), 6.74 (t, 1H), 2.89 (dt, 2H), 2.27 (t, 2H), 1.56 (m, 2H), 1.38 (m, 2H), 1.35 (s, 9H), 1.25 (m, 2H).

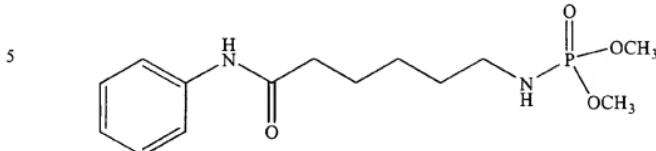
6-Aminohexanoic acid phenylamide, TFA salt (74)

25



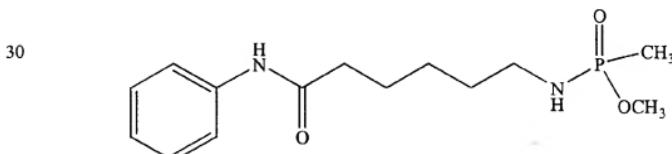
30 To a solution of carbamate 73 (300 mg, 0.98 mmol) in CH_2Cl_2 (15 mL) was added TFA (0.75 mL) and the solution stirred overnight. Complete consumption of starting material was confirmed by TLC. The mixture was evaporated under reduced pressure to remove all volatiles, leaving an off-white solid (295 mg, 0.92 mmol, 94%). 35 Crude 74 was used without further purification.

N-(N-Phenylcarbamoyl-5-pentyl)phosphoramidic acid dimethyl ester (75)



10 To a stirred suspension of ammonium salt **74** (197 mg, 0.62 mmol) and DIEA (148 μ L, 0.85 mmol) in CH_2Cl_2 (7 mL) at 0 $^{\circ}\text{C}$ was added dropwise dimethyl chlorophosphate (77 μ L, 0.72 mmol). The mixture was allowed to warm to ambient temperature and stirred overnight. The solution was diluted with H_2O (10 mL) and the 15 layers separated. The aqueous phase was extracted with CH_2Cl_2 (3 x 10 mL), the organic portions combined, washed with sat. NH_4Cl (5 mL), then brine, dried over MgSO_4 , and filtered. After concentration, the residue was purified by flash chromatography (2-5% MeOH/ CH_2Cl_2), and the fractions containing the more polar 20 of the two UV-active bands on TLC were combined and concentrated, giving **75** as a clear oil (40 mg, 0.13 mmol, 20%). TLC R_f 0.23 (5% MeOH/ CH_2Cl_2); $^1\text{H-NMR}$ (400 MHz, DMSO- d_6) δ 9.84 (s, 1H), 7.57 (d, 2H), 7.26 (t, 2H), 7.00 (t, 1H), 4.90 (dt, 1H), 3.51 (d, 6H), 2.71 (m, 2H), 2.28 (t, 2H), 1.56 (m, 2H), 1.40 (m, 2H), 1.29 (m, 2H).

Methyl *N*-(5-*N*-phenylcarbamoylpentyl)methylphosphonamidate (76)



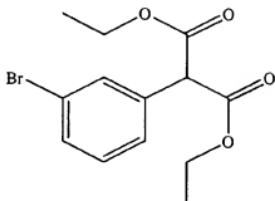
35 To a suspension of ammonium salt **74** (155 mg, 0.48 mmol) in CH_3CN (8 mL) were added DIEA (0.21 mL) and methyl

methylphosphonochloride (77 mg, 0.600 mmol). The reaction mixture was stirred overnight, during which time it clarified. The solution was partitioned between H₂O (10 mL) and EtOAc (15 mL) and the layers separated. The aqueous portion was extracted 5 with EtOAc (3 x 10 mL) and the organics combined, washed with sat. NH₄Cl (1 x 5 mL), then brine, dried over MgSO₄, and filtered. The product was purified by flash chromatography (3-10% MeOH/CH₂Cl₂), and the fractions containing the more polar spot were combined and concentrated to give 76 as a clear oil 10 (102 mg, 0.34 mmol, 71%). TLC R_f 0.16 (5% MeOH/CH₂Cl₂); ¹H-NMR (400 MHz, DMSO-d₆) δ 9.85 (s, 1H), 7.57 (d, 2H), 7.26 (t, 2H), 7.00 (t, 1H), 4.52 (dt, 1H), 3.43 (d, 3H), 2.73 (m, 2H), 2.28 (t, 2H), 1.57 (m, 2H), 1.38 (m, 2H), 1.28 (m, 2H), 1.26 (d, 3H).

15 Example 18 - Synthesis of Compound 77

Diethyl 3-bromophenylmalonate

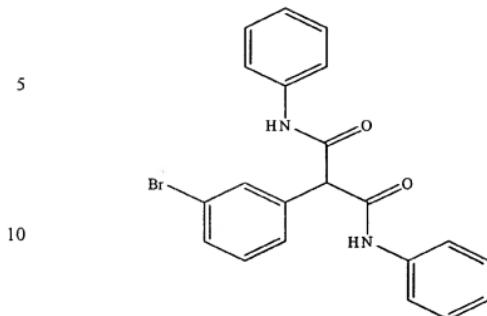
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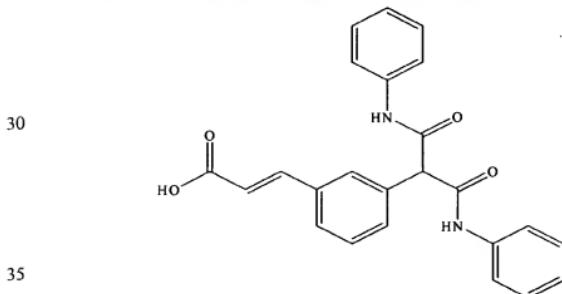
Diethyl 3-bromophenyl malonate was prepared according to the procedures of Cehnevert, R. and Desjardins, M. *Can. J. Chem.* 1994, 72, 3212-2317. ¹H NMR (CDCl₃, 300 MHz) δ 7.6 (s, 1H), 7.50 (d, 1H, J= 7.9 Hz), 7.37 (d, 1H, J=7.9 Hz), 7.26 (t, 1H, J=7.9 Hz), 4.58 (s, 1H), 4.22 (m, 4H), 1.29 (t, J=10 Hz).

3-bromophenyl malonyl di(phenylamide)



Diethyl 3-bromophenyl malonate (1 g, 3.2 mmol) was added to 15 aniline (5 mL). The reaction mixture was purged with Ar (g) and brought to reflux for 2h. After cooling, the reaction mixture was diluted with 10% HCl (20 mL) and ethyl acetate (50 mL). The organic layer was separated and concentrated to afford 3-bromophenyl malonyl di(phenylamide) as a white powder. (540 mg. 20 1.3 mmol, 42%). ¹H NMR (d6-DMSO, 300 MHz) δ 10.3 (bs, 2H), 7.65 (s, 1H), 7.60 (d, 4H, J=7.9 Hz), 7.54 (d, 1H, J=7.9 Hz), 7.46 (d, 1H, J=7.8 Hz), 7.35 (t, 1H, J=7.8 Hz), 7.31 (t, 4H, J=7.8 Hz), 7.06 (t, 2H, J=7.6 Hz), 4.91 (s, 1H).

25 3-(malonyl di(phenylamide)) cinnamic acid



3-bromophenyl malonyl di(phenylamide) (500 mg, 1.22 mmol),

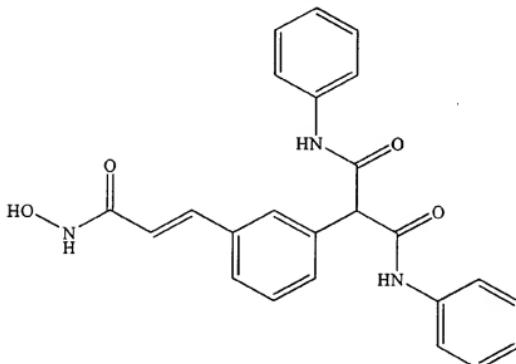
acrylic acid (115 mg, 1.6 mmol, 1.3 equiv.), $\text{Pd}(\text{OAc})_2$ (2 mg), tri-*o*-tolyl phosphine (20 mg), tributyl amine (0.6 mL) and xylenes (5 mL) were heated to 120°C for 6 h in a sealed vessel. After cooling, the reaction was diluted with 5% HCl (10 mL) and 5 ethyl acetate (50 mL). The organic layer was separated, filtered and on standing 3-(malonyl di(phenylamide)) cinnamic acid precipitated as a white powder (450 mg, 1.12 mmol, 92%). ^1H NMR (d6-DMSO, 300MHz, δ 12.4 (bs, 1H), 10.3 (bs, 2H), 7.73 (s, 1H), 7.7-7.5 (m, 6H), 7.52 (d, 1H, J =7.7 Hz), 7.43 (t, 1H, J =7.6 Hz), 7.31 (t, 4H, J =7.5Hz), 7.06 (t, 2H, J =7.4 Hz), 6.52 (d, 1H, J =16 Hz), 4.95 (s, 1H). APCI-MS 401 (M+1).

3-(malonyl di(phenylamide)) cinnamyl hydroxamic acid (77)

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3-(malonyl di(phenylamide)) cinnamic acid (200 mg, 0.5 mmol) was dissolved in dry CH_2Cl_2 (10mL). Isobutylchloroformate (0.10 mL, 0.77 mmol) and triethyl amine (0.20 mL) were added at 0°C with 30 stirring. After 2h at 25°C, O-(*t*-butyldiphenyl silyl)hydroxylamine was added and the mixture was stirred an additional 4h. The crude reaction mixture was applied directly to a pad a silica gel (15 g) and elution with 20% ethyl acetate/hexanes afforded the corresponding silyl protected 35 hydroxamic acid (R_f = 0.58, 50% ethyl acetate/hexanes) as a

foam. This was treated directly with 10% trifluoracetic acid in dichloromethane (10mL) for 4h. The solvents were concentrated at 50°C by rotavap and the residue was suspended in ethyl ether (10mL). Filtration of the resultant precipitate 5 afforded compound 77 as a white powder (150 mg, 0.365 mmol, 73%). ¹H NMR (d6-DMSO, 300 MHz, δ 10.8 (bs, 0.5H), 10.2 (bs, 2H), 9.06 (bs, 0.5H), 7.7-7.55 (m, 5H), 7.53-7.38 (m, 4H), 7.31 (t, 4H, J=7.7 Hz), 7.06 (t, 2H, J=7.3 Hz), 6.50 (d, 1H, J=16Hz), 4.92 (s, 1H). APCI-MS 416 (M+1).

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The effect of compound 77 on MEL cell differentiation and Histone Deacetylase activity is shown in Table 2. Compound 77 corresponds to structure 683 in Table 2. As evident from Table 2, compound 77 is expected to be a highly effective 15 cytodifferentiating agent.

Results
All the compounds which were prepared were tested. Table 2 below shows the results of testing of only a subgroup of 20 compounds. Table 2 is compiled from experiments similar to the experiments described in Examples 7-10 above. The tested compounds were assigned structure numbers as shown in Table 2. The structure numbers were randomly assigned and do not correlate to the compound numbers used elsewhere in this 25 disclosure.

The results shown in Table 2 verify the general accuracy of the predictive principals for the design of compounds having cell differentiation and HDAC inhibition activity discussed above in 30 this disclosure. Based on the principals and synthesis schemes disclosed, a number of additional compounds can readily be designed, prepared and tested for cell differentiation and HDAC inhibition activity.

35 Figures 11a-f show the effect of selected compounds on affinity

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purified human epitope-tagged (Flag) HDAC1. The effect was assayed by incubating the enzyme preparation in the absence of substrate on ice for 20 minutes with the indicated amounts of compound. Substrate([³H]acetyl-labeled murine erythroleukemia 5 cell-derived histones) was added and the samples were incubated for 20 minutes at 37°C in a total volume of 30 μ l. The reactions were then stopped and released acetate was extracted and the amount of radioactivity released determined by scintillation counting. This is a modification of the HDAC 10 Assay described in Richon et al. 1998 (39).

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Table 2 - Inhibition data of selected compounds.

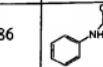
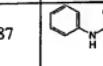
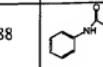
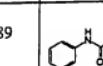
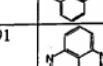
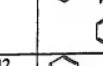
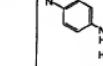
NO:	Structure	MEL Diff				HDAC inh	
		Range	Opt.	% B+	cells/ml $\times 10^5$	Range	ID50
SAHA (390)		0.5 to 50 μ M	2.5 μ M	68	3.6	0.001 to 100 μ M	200 nM
654		0.1 to 50 μ M	200 nM	44	9	0.0001 to 100 μ M	1 nM
655		0.1 to 50 μ M	400 nM	16	3.3	0.01 to 100 μ M	100 nM
656		0.4 to 50 μ M		0		0.01 to 100 μ M	>100 μ M
657		0.4 to 50 μ M		0		0.01 to 100 μ M	>100 μ M
658		0.01 to 50 μ M	40 nM	8	13	0.0001 to 100 μ M	2.5 nM
659		0.4 to 50 μ M		0		0.01 to 100 μ M	10 μ M
660		0.2 to 12.5 μ M	800 nM	27		0.001 to 100 μ M	50 nM
661		0.1 to 50 μ M	500 nM	7		0.01 to 100 μ M	20 nM
662		0.2 to 50 μ M		0		0.001 to 100 μ M	>100 μ M

No.	Structure	MEL Cell Differentiation				HDAC1 Inhibition	
		Range	Opt.	%B+	cells/ ml x 10 ⁻⁵	Range	ID50
663		0.2 to 50 μ M	200 nM	43	7	0.001 to 100 μ M	100 nM
664		0.2 to 50 μ M	400 nM	33	22	0.001 to 100 μ M	50 nM
665		0.1 – 50 μ M	150 nM	24	30	0.001 to 100 μ M	50 nM
666		0.1 – 50 μ M	150 nM	31	28	0.001 to 100 μ M	100 nM
667		0.02 – 10 μ M	80 nM	27	2	0.001 to 100 μ M	50 nM
668		0.02 to 10 μ M	10 μ M	11	4.7	0.001 to 100 μ M	100 nM
669		0.8 to 50 μ M	4 μ M	11	16.0	0.001 to 100 μ M	10 μ M

No.	Structure	MEL Cell Differentiation				HDAC1 Inhibition	
		Range	Opt.	%B+	Cells/ml x 10 ³	Range	ID50
670		0.4 to 50 μ M	No effect up to 25 μ M	-	13.0	0.001 to 100 μ M	>100 μ M
671		0.4 to 50 μ M	3.1 μ M	35	12.5	0.001 to 100 μ M	200 nM
672		0.8 to 50 μ M		0	No Inh	0.01 to 100 μ M	100 μ M
673		0.8 to 50 μ M		0	No Inh	0.01 to 100 μ M	100 μ M
674		0.8 to 50 μ M		0	Dead at 25 μ M	0.01 to 100 μ M	50 μ M
675		0.8 to 50 μ M		0	No Inh	0.001 to 100 μ M	>100 μ M
676		0.8 to 50 μ M		0	No Inh	0.01 to 100 μ M	100 μ M
677		0.05 to 25 μ M	1.6 μ M	23	4.5	0.001 to 100 μ M	5 nM

No.	Structure	MEL cell differentiation				HDAC Inh	
		Range	Opt.	%B+	cells/ml x10-5	Range	ID50
678		0.8 to 50 μ M		0	No Inh	0.001 to 100 μ M	>100 μ M
679		0.8 to 50 μ M		0	No inh	0.001 to 100 μ M	>100 μ M
680						0.01 to 100 μ M	>100 μ M

No.	Structure	MEL cell differentiation				HDAC Inh	
		Range	Opt.	%B+	cells/ml x10-5	Range	ID50
681		0.8 to 50 μ M	3 μ M	3	2.5	0.01 to 100 μ M	200 nM
682		0.8 to 50 μ M	50 μ M	8	1.1	0.01 to 100 μ M	150 nM
683		0.01 to 0.1 μ M	20 nM	9	9.0	0.0001 to 100 μ M	1 nM
684		0.4 to 50 μ M		0	No inh	0.01 to 100 μ M	100 μ M
685		0.125 to 5 μ M	1.0 μ M	20	1.0	0.01 to 100 μ M	150 nM

No.	Structure	MEL cell differentiation				HDAC Inh	
		Range	Opt.	%B+	cells/ml x10-5	Range	ID50
686		0.4 to 50 μ M		0	No inh	0.01 to 100 μ M	100 μ M
687		0.125 to 5 μ M		0	No inh	0.01 to 100 μ M	200 nM
688		0.4 to 50 μ M		0	No inh	0.01 to 100 μ M	>100 μ M
689		5.0 to 40 μ M	35 μ M	48	2.0	0.01 to 100 μ M	200 nM
690		5.0 to 40 μ M	10 μ M	38	2.5	0.01 to 100 μ M	150 nM
691		1.0 to 25 μ M		0	No inh	0.01 to 100 μ M	100 nM
692		0.03 to 5 μ M	1 μ M	27	18.0	0.01 to 100 μ M	1 nM
693		0.4 to 50 μ M		0	No inh	0.01 to 100 μ M	>100 μ M

BIBLIOGRAPHY

1. Sporn, M. B., Roberts, A. B., and Driscoll, J. S. (1985) in *Cancer: Principles and Practice of Oncology*, eds. Hellman, S., Rosenberg, S. A., and DeVita, V. T., Jr., Ed. 2, (J. B. Lippincott, Philadelphia), P. 49.
2. Breitman, T. R., Selonick, S. E., and Collins, S. J. (1980) *Proc. Natl. Acad. Sci. USA* 77: 2936-2940.
3. Olsson, I. L. and Breitman, T. R. (1982) *Cancer Res.* 42: 3924-3927.
4. Schwartz, E. L. and Sartorelli, A. C. (1982) *Cancer Res.* 42: 2651-2655.
5. Marks, P. A., Sheffery, M., and Rifkind, R. A. (1987) *Cancer Res.* 47: 659.
6. Sachs, L. (1978) *Nature (Lond.)* 274: 535.
7. Friend, C., Scher, W., Holland, J. W., and Sato, T. (1971) *Proc. Natl. Acad. Sci. (USA)* 68: 378-382.
8. Tanaka, M., Levy, J., Terada, M., Breslow, R., Rifkind, R. A., and Marks, P. A. (1975) *Proc. Natl. Acad. Sci. (USA)* 72: 1003-1006.
9. Reuben, R. C., Wife, R. L., Breslow, R., Rifkind, R. A., and Marks, P. A. (1976) *Proc. Natl. Acad. Sci. (USA)* 73: 862-866.
10. Abe, E., Miyaura, C., Sakagami, H., Takeda, M., Konno, K., Yamazaki, T., Yoshika, S., and Suda, T. (1981) *Proc. Natl. Acad. Sci. (USA)* 78: 4990-4994.

11. Schwartz, E. L., Snoddy, J. R., Kreutter, D., Rasmussen, H., and Sartorelli, A. C. (1983) Proc. Am. Assoc. Cancer Res. 24: 18.
12. Tanenaga, K., Hozumi, M., and Sakagami, Y. (1980) Cancer Res. 40: 914-919.
13. Lottem, J. and Sachs, L. (1975) Int. J. Cancer 15: 731-740.
14. Metcalf, D. (1985) Science, 229: 16-22.
15. Scher, W., Scher, B. M., and Waxman, S. (1983) Exp. Hematol. 11: 490-498.
16. Scher, W., Scher, B. M., and Waxman, S. (1982) Biochem. & Biophys. Res. Comm. 109: 348-354.
17. Huberman, E. and Callaham, M. F. (1979) Proc. Natl. Acad. Sci. (USA) 76: 1293-1297.
18. Lottem, J. and Sachs, L. (1979) Proc. Natl. Acad. Sci. (USA) 76: 5158-5162.
19. Terada, M., Epner, E., Nudel, U., Salmon, J., Fibach, E., Rifkind, R. A., and Marks, P. A. (1978) Proc. Natl. Acad. Sci. (USA) 75: 2795-2799.
20. Morin, M. J. and Sartorelli, A. C. (1984) Cancer Res. 44: 2807-2812.
21. Schwartz, E. L., Brown, B. J., Nierenberg, M., Marsh, J. C., and Sartorelli, A. C. (1983) Cancer Res. 43: 2725-2730.
22. Sugano, H., Furusawa, M., Kawaguchi, T., and Ikawa, Y.

-104-

- (1973) *Bibl. Hematol.* 39: 943-954.
23. Ebert, P. S., Wars, I., and Buell, D. N. (1976) *Cancer Res.* 36: 1809-1813.
24. Hayashi, M., Okabe, J., and Hozumi, M. (1979) *Gann* 70: 235-238.
25. Fibach, E., Reuben, R. C., Rifkind, R. A., and Marks, P. A. (1977) *Cancer Res.* 37: 440-444.
26. Melloni, E., Pontremoli, S., Damiani, G., Viotti, P., Weich, N., Rifkind, R. A., and Marks, P. A. (1988) *Proc. Natl. Acad. Sci. (USA)* 85: 3835-3839.
27. Reuben, R., Khanna, P. L., Gazitt, Y., Breslow, R., Rifkind, R. A., and Marks, P. A. (1978) *J. Biol. Chem.* 253: 4214-4218.
28. Marks, P. A. and Rifkind, R. A. (1988) *International Journal of Cell Cloning* 6: 230-240.
29. Melloni, E., Pontremoli, S., Michetti, M., Sacco, O., Cakiroglu, A. G., Jackson, J. F., Rifkind, R. A., and Marks, P. A. (1987) *Proc. Natl. Acad. Sciences (USA)* 84: 5282-5286.
30. Marks, P. A. and Rifkind, R. A. (1984) *Cancer* 54: 2766-2769.
31. Egorin, M. J., Sigman, L. M. VanEcho, D. A., Forrest, A., Whitacre, M. Y., and Aisner, J. (1987) *Cancer. Res.* 47: 617-623.
32. Rowinsky, E. W., Ettinger, D. S., Grochow, L. B.,

- Brundrett, R. B., Cates, A. E., and Donehower, R. C. (1986) *J. Clin. Oncol.* 4: 1835-1844.
33. Rowinsky, E. L. Ettinger, D. S., McGuire, W. P., Noe, D. A., Grochow, L. B., and Donehower, R. C. (1987) *Cancer Res.* 47: 5788-5795.
34. Callery, P. S., Egorin, M. J., Geelhaar, L. A., and Nayer, M. S. B. (1986) *Cancer Res.* 46: 4900-4903.
35. Young, C. W. Fanucchi, M. P., Walsh, T. B., Blatzer, L., Yaldaie, S., Stevens, Y. W., Gordon, C., Tong, W., Rifkind, R. A., and Marks, P. A. (1988) *Cancer Res.* 48: 7304-7309.
36. Andreeff, M., Young, C., Clarkson, B., Fetten, J., Rifkind, R. A., and Marks, P. A. (1988) *Blood* 72: 186a.
37. Marks, P.A., Richon, V.M., Breslow, R., Rifkind, R.A., *Life Sciences* 1999, 322: 161-165.
38. Yoshida et al., 1990, *J. Biol. Chem.* 265:17174-17179.
39. Richon, V.M., Emiliani, S., Verdin, E., Webb, Y., Breslow, R., Rifkind, R.A., and Marks, P.A., *Proc. Natl. Acad. Sci. (USA)* 95: 3003-3007 (1998).
40. Nishino, N. et. al. *Chem. Pharm. Bull.* 1996, 44, 212-214.
41. U.S. Patent No. 5,369,108, issued November 29, 1994.
42. Kijima et al., 1993, *J. Biol. Chem.* 268:22429-22435.
43. Lea et al., 1999, *Int. J. Oncol.* 2:347-352.
44. Kim et al., 1999, *Oncogene* 15:2461-2470.

45. Saito et al., 1999, *Proc. Natl. Acad. Sci.* 96:4592-4597.
46. Lea and Tulssyan, 1995, *Anticancer Res.* 15:879-883.
47. Nokajima et al., 1998, *Exp. Cell Res.* 241:126-133.
48. Kwon et al., 1998, *Proc. Natl. Acad. Sci. USA* 95:3356-3361.
49. Richon et al, 1996, *Proc. Natl. Acad. Sci. USA* 93:5705-5708.
50. Kim et al., 1999, *Oncogene* 18:2461-2470.
51. Yoshida et al., 1995, *Bioessays* 17:423-430.
52. Yoshida & Beppu, 1988, *Exp. Cell. Res.* 177:122-131.
53. Warrell et al., 1998, *J. Natl. Cancer Inst.* 90:1621-1625.
54. Desai et al., 1999, *Proc. AACR* 40: abstract #2396.
55. Cohen et al., *Antitumor Res.*, submitted.
56. D. W. Christianson and W. N. Lipscomb, "The Complex Between Carboxypeptidase A and a Possible Transition-State Analogue: Mechanistic Inferences from High-Resolution X-ray Structures of Enzyme-inhibitor Complexes," *J. Am. Chem. Soc.* 1986, 108, 4998-5003.
57. G. H. S. Prakash and A. K. Yudin, "Perfluoroalkylation with Organosilicon Reagents," *Chem. Rev.* 1997, 97, 757-786.
58. J.-C. Blazejewski, E. Anselmi, and M. P. Wilmshurst, "Extending the Scope of Ruppert's Reagent:

- Trifluoromethylation of Imines," *Tet. Letters* **1999**, 40, 5475-5478.
59. R. J. Linderman and D. M. Graves, "Oxidation of Fluoroalkyl-Substituted Carbinols by the Dess-Martin Reagent," *J. Org. Chem.* **1989**, 54, 661-668.
60. N. E. Jacobsen and P. A. Bartlett, "A Phosphonamide Dipeptide Analogue as an Inhibitor of Carboxypeptidase A," *J. Am. Chem. Soc.* **1981**, 103, 654-657.
61. S. Lindskog, L. E. Henderson, K. K. Kannan, A. Liljas, P. O. Nyman, and B. Strandberg, "Carbonic Anhydrase", in *The Enzymes*, 3rd edition, P.D. Boyer, ed., **1971**, vol. V, pp. 587-665, see p. 657.
62. Durrant, G.; Greene, R.H.; Lambeth, P.F.; Lester, M.G.; Taylor, N.R., *J. Chem. Soc., Perkin Trans. I* **1983**, 2211-2214.
63. Burden, R.S.; Crombie, L., *J. Chem. Soc. (C)* **1969**, 2477-2481.
64. Farquhar, D.; Cherif, A.; Bakina, E.; Nelson, J.A., *J. Med. Chem.*, **1998**, 41, 965-972.
65. Boivin, J.; El Kaim, L.; Zard, S.Z., *Tet. Lett.* **1992**, 33, 1285-1288.
66. Finnin, M.S. et al., Structures of a histone deacetylase homologue bound to the TSA and SAHA inhibitors. *Nature* **401**, 188-93 (1999).
67. Webb, Y. et al., Photoaffinity labeling and mass

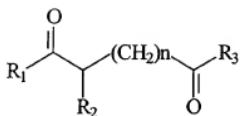
-108-

spectrometry identify ribosomal protein S3 as a potential target for hybrid polar cytodifferentiation agents. *J. Biol. Chem.* **274**, 14280-14287 (1997).

68. Butler, L.M. et al., Suberoylanilide hydroxamic acid (SAHA), an inhibitor of histone deacetylase, suppresses the growth of the CWR22 human prostate cancer xenograft. *submitted* (2000).

What is claimed is:

1. A compound having the formula:



wherein R₁ and R₂ are the same or different and are each a hydrophobic moiety;

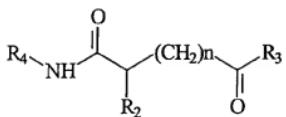
wherein R₃ is a hydroxamic acid, hydroxylamino, hydroxyl, amino, alkylamino, or alkyloxy group; and

n is an integer from 3 to 10,

or a pharmaceutically acceptable salt thereof.

2. The compound of claim 1, wherein each of R₁ and R₂ is directly attached or through a linker, and is, substituted or unsubstituted, aryl, cycloalkyl, cycloalkylamino, naphtha, pyridineamino, piperidino, 9-purine-6-amine, thiazoleamino group, hydroxyl, branched or unbranched alkyl, alkenyl, alkyloxy, aryloxy, arylalkyloxy, or pyridine group.
3. The compound of claim 2 wherein the linker is an amide moiety, -O-, -S-, -NH-, or -CH₂-.

4. The compound of claim 1 having the formula:

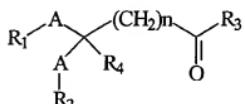


wherein each of R_4 is, substituted or unsubstituted, aryl, cycloalkyl, cycloalkylamino, naphtha, pyridineamino, piperidino, 9-purine-6-amine, thiazoleamino group, hydroxyl, branched or unbranched alkyl, alkenyl, alkyloxy, aryloxy, arylalkyloxy, or pyridine group.

5. The compound of claim 4, wherein R_2 is -amide- R_5 ,

wherein R_5 is, substituted or unsubstituted, aryl, cycloalkyl, cycloalkylamino, naphtha, pyridineamino, piperidino, 9-purine-6-amine, thiazoleamino group, hydroxyl, branched or unbranched alkyl, alkenyl, alkyloxy, aryloxy, arylalkyloxy, or pyridine group.

6. A compound having the formula:



wherein each of R_1 and R_2 is, substituted or unsubstituted, aryl, cycloalkyl, cycloalkylamino, naphtha, pyridineamino, piperidino, 9-purine-6-amine, thiazoleamino group, hydroxyl, branched or unbranched alkyl, alkenyl, alkyloxy, aryloxy, arylalkyloxy, or pyridine group;

wherein R_3 is a hydroxamic acid, hydroxylamino, hydroxyl,

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amino, alkylamino, or alkyloxy group;

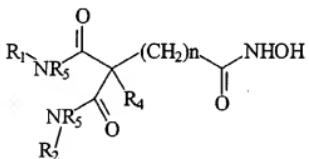
wherein R₄ is hydrogen, a halogen, a phenyl, or a cycloalkyl moiety;

wherein A may be the same or different and represents an amide moiety, -O-, -S-, -NR₅-, or -CH₂-, where R₅ is a substituted or unsubstituted C₁-C₅ alkyl; and

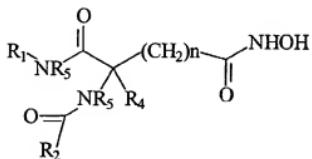
wherein n is an integer from 3 to 10,

or a pharmaceutically acceptable salt thereof.

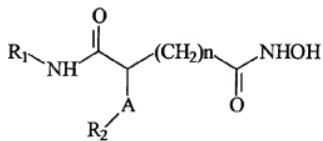
7. The compound of claim 6 having the formula:



8. The compound of claim 6 having the formula:



9. The compound of claim 6 having the formula:

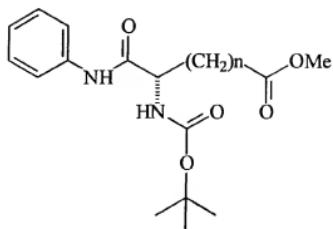


wherein each of R_1 and R_2 is, substituted or unsubstituted, aryl, cycloalkyl, cycloalkylamino, naphtha, pyridineamino, piperidino, *t*-butyl, aryloxy, arylalkyloxy, or pyridine group; and

wherein n is an integer from 3 to 8.

10. The compound of claim 9 wherein the aryl or cycloalkyl group is substituted with a methyl, cyano, nitro, trifluoromethyl, amino, aminocarbonyl, methylcyano, chloro, fluoro, bromo, iodo, 2,3-difluoro, 2,4-difluoro, 2,5-difluoro, 3,4-difluoro, 3,5-difluoro, 2,6-difluoro, 1,2,3-trifluoro, 2,3,6-trifluoro, 2,4,6-trifluoro, 3,4,5-trifluoro, 2,3,5,6-tetrafluoro, 2,3,4,5,6-pentafluoro, azido, hexyl, *t*-butyl, phenyl, carboxyl, hydroxyl, methoxy, phenyloxy, benzylxy, phenylaminoxy, phenylaminocarbonyl, methyoxy carbonyl, methylaminocarbonyl, dimethylamino, dimethylaminocarbonyl, or hydroxylaminocarbonyl group.

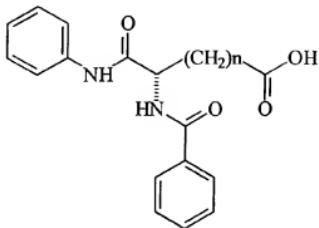
11. The compound of claim 6 having the formula:



or an enantiomer thereof.

12. The compound of claim 11, wherein n=5.

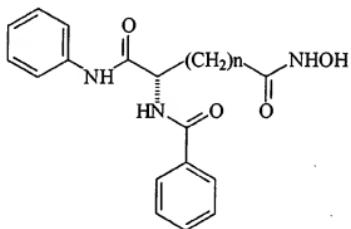
13. The compound of claim 6 having the formula:



or an enantiomer thereof.

14. The compound of claim 13, wherein n=5.

15. The compound of claim 6 having the formula:

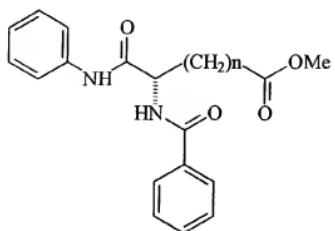


or an enantiomer thereof.

16. The compound of claim 15, wherein n=5.

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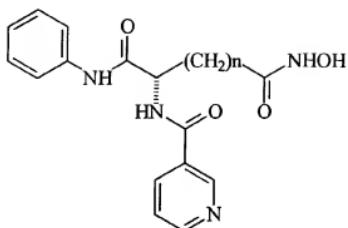
17. The compound of claim 6 having the formula:



or an enantiomer thereof.

18. The compound of claim 17, wherein n=5.

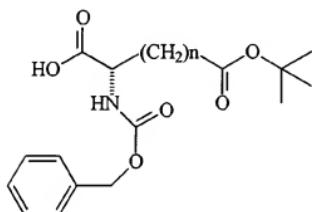
19. The compound of claim 6 having the formula:



or an enantiomer thereof.

20. The compound of claim 19, wherein n=5.

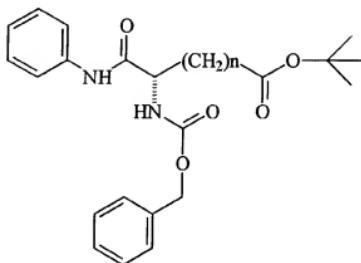
21. The compound of claim 6 having the formula:



or an enantiomer thereof.

22. The compound of claim 21, wherein n=5.

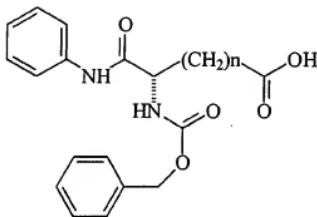
23. The compound of claim 6 having the formula:



or an enantiomer thereof.

24. The compound of claim 23, wherein n=5.

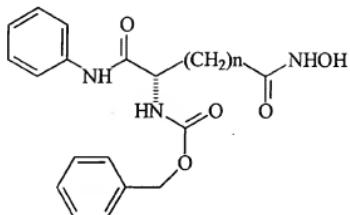
25. The compound of claim 6 having the formula:



or an enantiomer thereof.

26. The compound of claim 25, wherein n=5.

27. The compound of claim 6 having the formula:

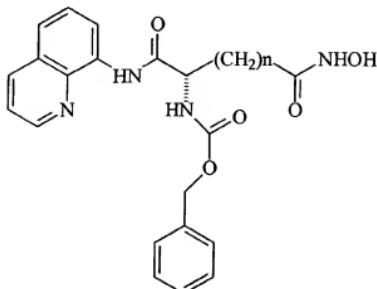


or an enantiomer thereof.

28. The compound of claim 27, wherein n=5.

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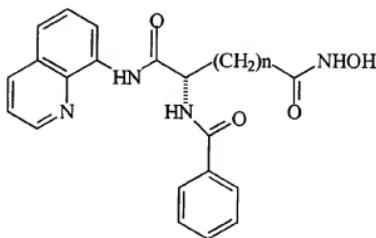
29. The compound of claim 6 having the formula:



or an enantiomer thereof.

30. The compound of claim 29, wherein n=5.

31. The compound of claim 6 having the formula:

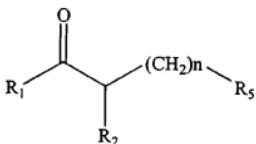


or an enantiomer thereof.

32. The compound of claim 31, wherein n=5.

33. A pharmaceutical composition comprising a pharmaceutically effective amount of the compound of any one of claims 1-9 and a pharmaceutically acceptable carrier.

34. A method of selectively inducing terminal differentiation of neoplastic cells and thereby inhibiting proliferation of such cells which comprises contacting the cells under suitable conditions with an effective amount of the compound of any one of claims 1-9.
35. A method of treating a patient having a tumor characterized by proliferation of neoplastic cells which comprises administering to the patient an effective amount of the compound of any one of claims 1-9.
36. A compound having the formula:



wherein R₁ and R₂ are the same or different and are each a hydrophobic moiety;

wherein R₅ is -C(O)-NHOH (hydroxamic acid), -C(O)-CF₃ (trifluoroacetyl), -NH-P(O)OH-CH₃, -SO₂NH₂ (sulfonamide), -SH (thiol), -C(O)-R₆, wherein R₆ is hydroxyl, amino, alkylamino, or alkyloxy group; and

n is an integer from 3 to 10,

or a pharmaceutically acceptable salt thereof.

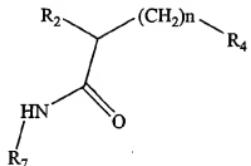
37. The compound of claim 36, wherein each of R₁ and R₂ is directly attached or through a linker, and is, substituted or unsubstituted, aryl, cycloalkyl, cycloalkylamino, naphtha, pyridineamino, piperidino, 9-purine-6-amine,

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thiazoleamino group, hydroxyl, branched or unbranched alkyl, alkenyl, alkyloxy, aryloxy, arylalkyloxy, or pyridine group.

38. The compound of claim 37, wherein the linker is an amide moiety, -O-, -S-, -NH-, or -CH₂-.

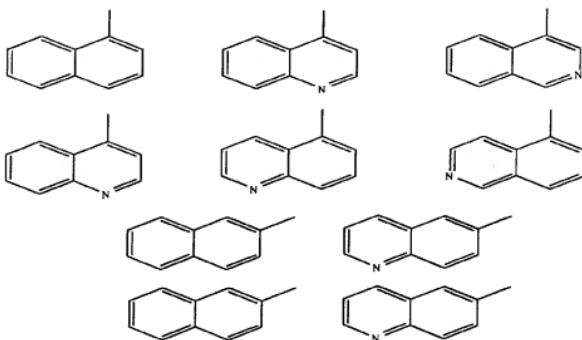
39. The compound of claim 36 having the formula:



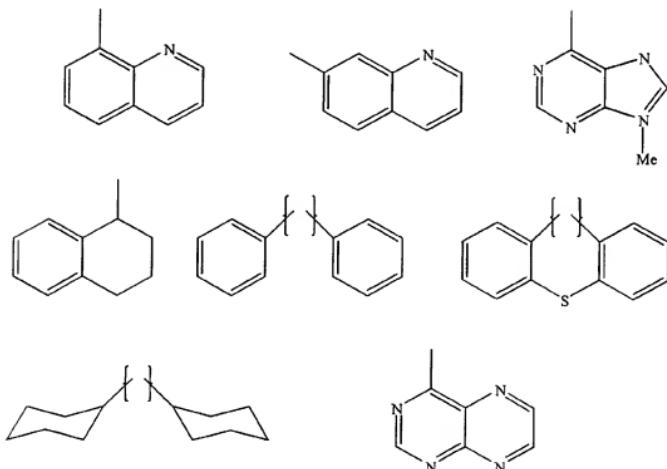
wherein each of R₁ is, substituted or unsubstituted, aryl, cycloalkyl, cycloalkylamino, naphtha, pyridineamino, piperidino, 9-purine-6-amine, thiazoleamino group, hydroxyl, branched or unbranched alkyl, alkenyl, alkyloxy, aryloxy, arylalkyloxy, or pyridine group.

40. The compound of claim 39, wherein R₂ is -sulfonamide-R₈, or -amide-R₈, wherein R₈ is, substituted or unsubstituted, aryl, cycloalkyl, cycloalkylamino, naphtha, pyridineamino, piperidino, 9-purine-6-amine, thiazoleamino group, hydroxyl, branched or unbranched alkyl, alkenyl, alkyloxy, aryloxy, arylalkyloxy, or pyridine group.

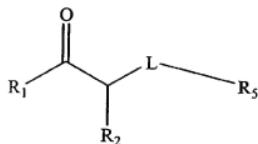
41. The compound of claim 39, wherein R₂ is -NH-C(O)-Y, -NH-SO₂-Y, wherein Y is selected from the group consisting of:



42. The compound of claim 39, wherein R₇ is selected from the group consisting of:



43. A compound having the formula:



wherein R₁ and R₂ are the same or different and are each a hydrophobic moiety;

wherein R₅ is -C(O)-NHOH (hydroxamic acid), -C(O)-CF₃ (trifluoroacetyl), -NH-P(O)OH-CH₃, -SO₂NH₂ (sulfonamide), -SH (thiol), -C(O)-R₆, wherein R₆ is hydroxyl, amino, alkylamino, or alkyloxy group; and

wherein L is a linker consisting of -(CH₂)-, -C(O)-, -S-, -O-, -(CH=CH)-, -phenyl-, or -cycloalkyl-, or any combination thereof,

or a pharmaceutically acceptable salt thereof.

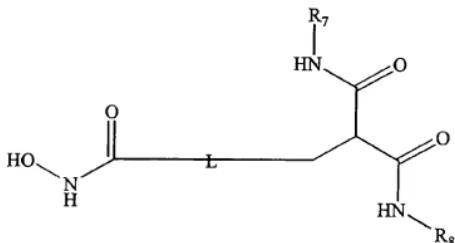
44. The compound of claim 43, wherein n is from 4-7, and m is from 1-3.

45. The compound of claim 43, wherein each of R₁ and R₂ is directly attached or through a linker, and is, substituted or unsubstituted, aryl, cycloalkyl, cycloalkylamino, naphtha, pyridineamino, piperidino, 9-purine-6-amine, thiazoleamino group, hydroxyl, branched or unbranched alkyl, alkenyl, alkyloxy, aryloxy, arylalkyloxy, or pyridine group.

46. The compound of claim 43, wherein the linker is an amide moiety, -O-, -S-, -NH-, or -CH₂-.

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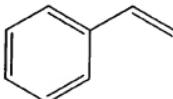
47. The compound of claim 43, having the formula:



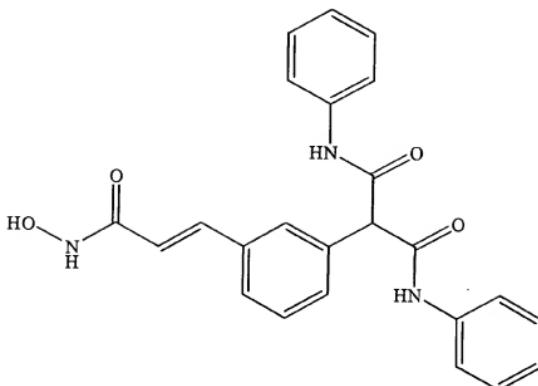
wherein L is a linker selected from the group consisting of $-(\text{CH}_2)-$, $-(\text{CH}=\text{CH})-$, -phenyl-, -cycloalkyl-, or any combination thereof; and

wherein each of R_7 and R_8 are independently substituted or unsubstituted, aryl, cycloalkyl, cycloalkylamino, naphtha, pyridineamino, piperidino, 9-purine-6-amine, thiazoleamino group, hydroxyl, branched or unbranched alkyl, alkenyl, alkyloxy, aryloxy, arylalkyloxy, or pyridine group.

48. The compound of claim 47, wherein the linker L comprises the moiety

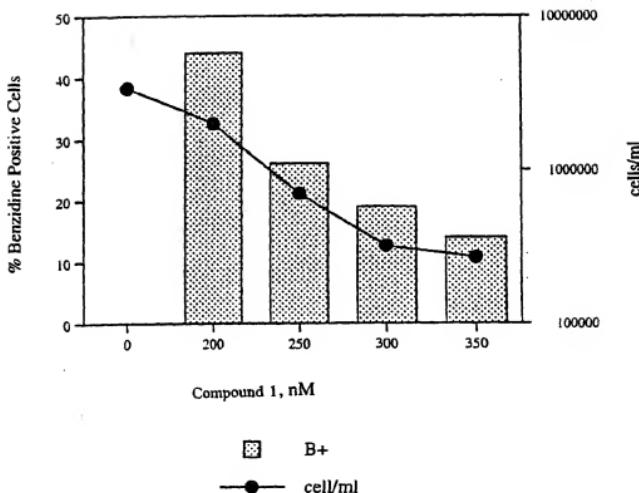


49. The compound of claim 43, having the formula:



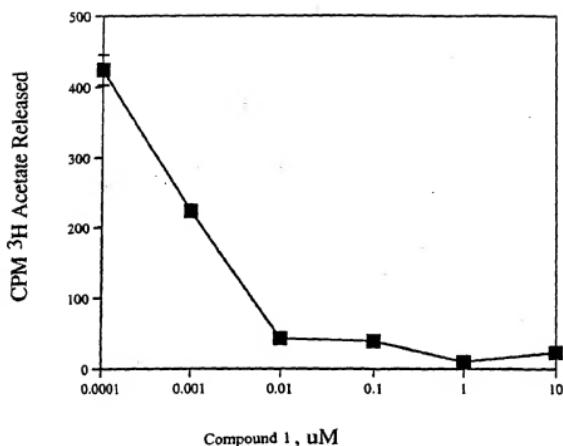
50. A pharmaceutical composition comprising the compound of claim 1, 36 or 43 and a pharmaceutically acceptable carrier.
51. A pharmaceutically acceptable salt of the compound of claim 1, 36, or 43.
52. A prodrug of the compound of claim 1, 36 or 43.
53. A method of inducing differentiation of tumor cells in a tumor comprising contacting the cells with an effective amount of the compound of claim 1, 36 or 43 so as to thereby differentiate the tumor cells.
54. A method of inhibiting the activity of histone deacetylase comprising contacting the histone deacetylase with an effective amount of the compound of claim 1, 36 or 43 so as to thereby inhibit the activity of histone deacetylase.

Figure 1



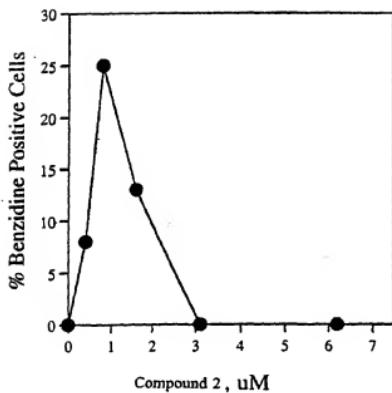
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Figure 2



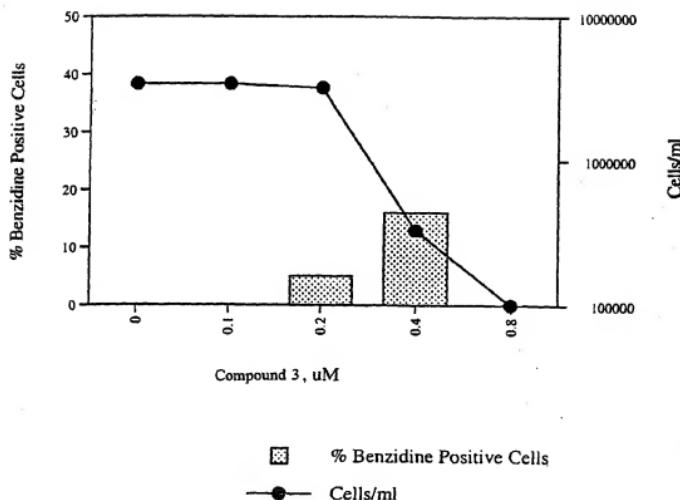
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Figure 3



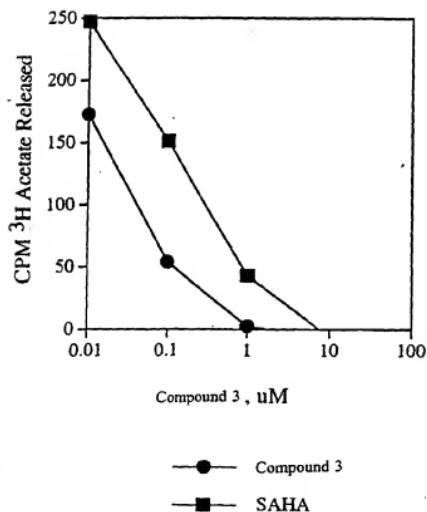
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Figure 4



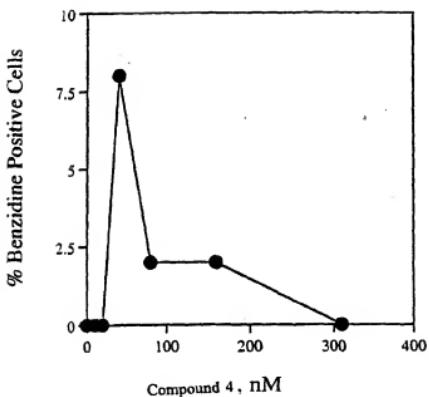
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Figure 5



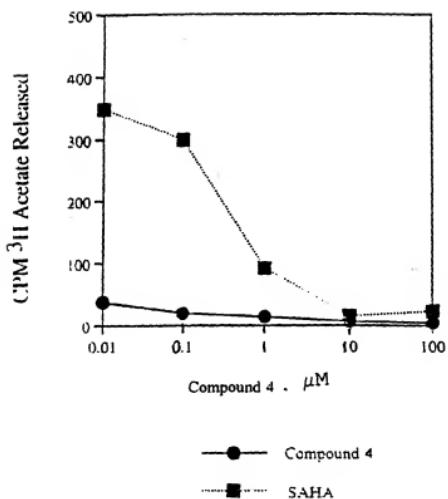
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Figure 6



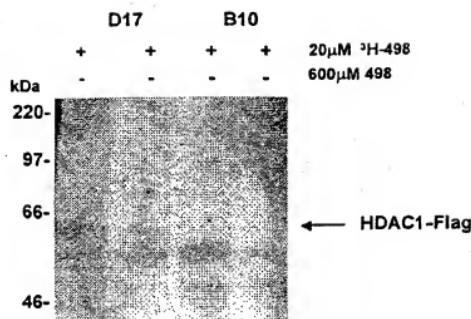
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Figure 7



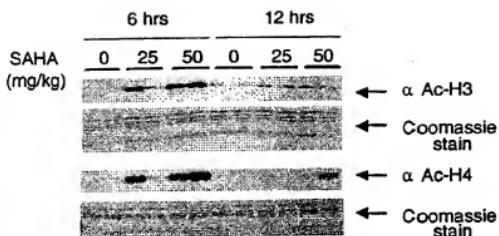
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Figure 8



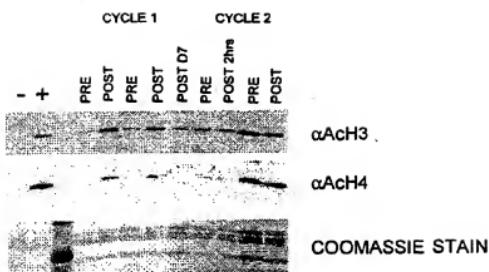
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Figure 9



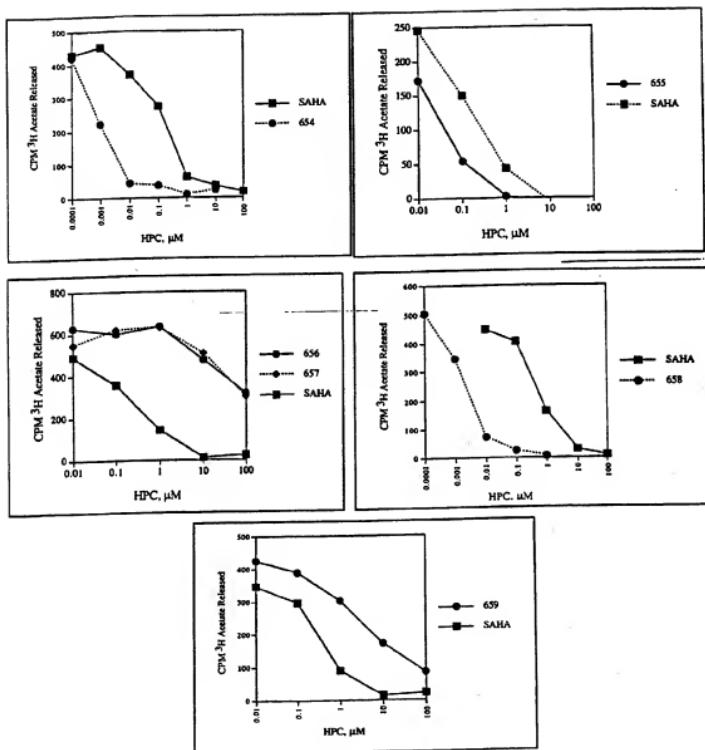
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Figure 10



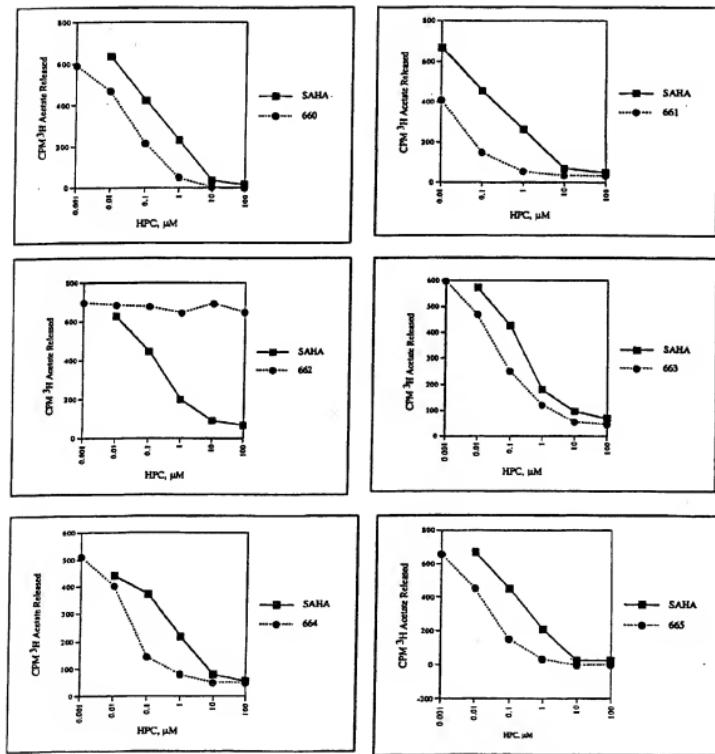
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Fig. 11a



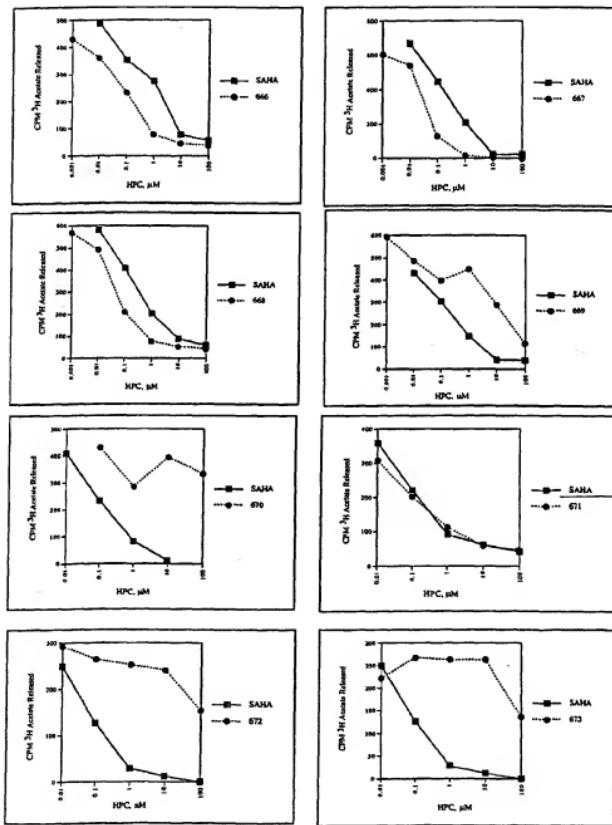
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Fig. 11b

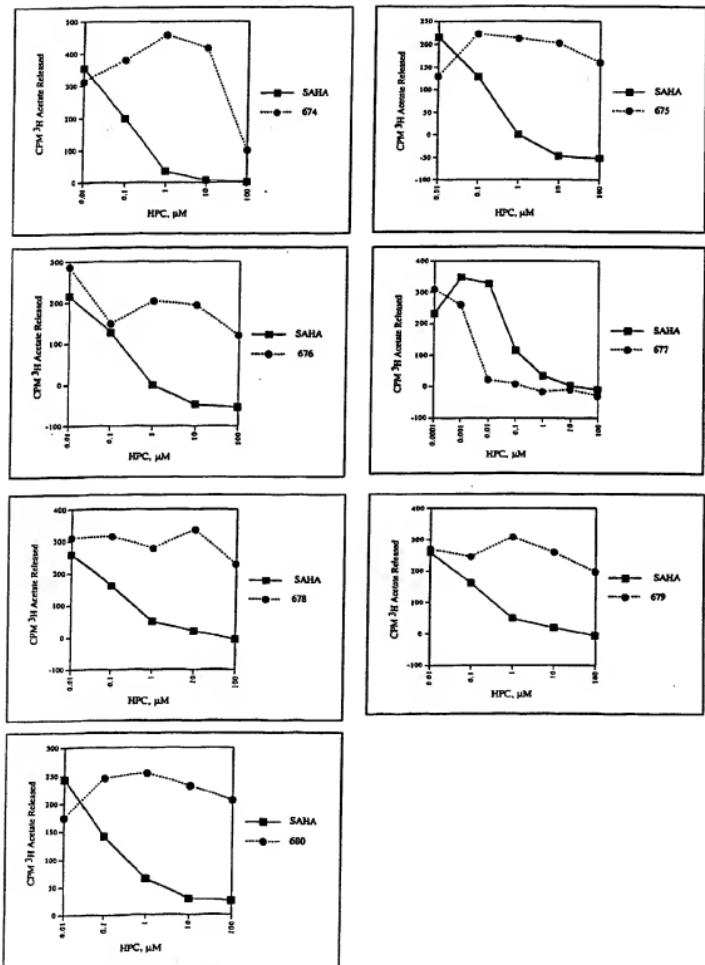


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Fig. 11c

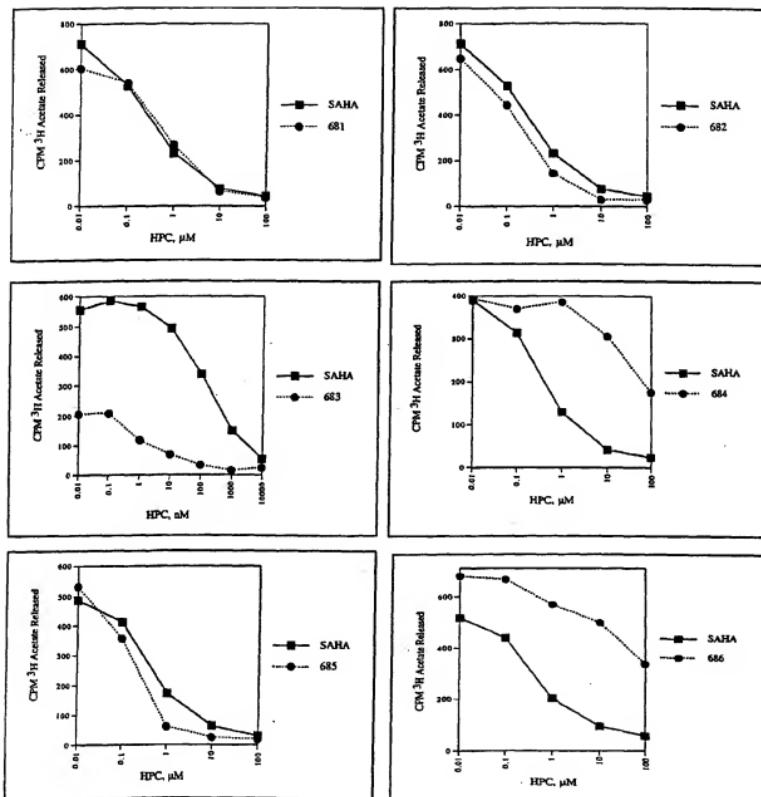


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Fig. 11d



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Fig. 11e



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Fig. 11f

